



**Full Length Article**

## Comparison of Absorption Characteristics of Iron Glycine Chelate and Ferrous Sulfate in Caco-2 Cells

Wen-Qiang Ma<sup>1,2</sup>, Jing Wu<sup>1</sup>, Zhao Zhuo<sup>1</sup>, Hong Sun<sup>1</sup>, Min Yue<sup>1</sup> and Jie Feng<sup>1\*</sup>

<sup>1</sup>College of Animal Sciences, Zhejiang University, Hangzhou, 310058, P.R. China

<sup>1</sup>The Key Laboratory of Molecular Animal Nutrition, Ministry of Education, College of Animal Science, Zhejiang University, Hangzhou, 310029, P.R. China

<sup>2</sup>The Key Laboratory of Animal Physiology and Biochemistry, College of Veterinary Medicine, Nanjing Agricultural University, Nanjing, 210095, PR China

\*For correspondence: [fengj@zju.edu.cn](mailto:fengj@zju.edu.cn)

### Abstract

The study was conducted to compare the absorption characteristics of iron glycine chelate (Fe-Gly) and ferrous sulfate (FeSO<sub>4</sub>) in Caco-2 cells. Effects of several factors, including concentration, time and temperature on iron (Fe) transport were investigated. In Caco-2 cell model, Fe transport amount in Fe-Gly treatment was higher than that in FeSO<sub>4</sub> from the apical side (AP) to the basolateral (BL) side. However, there was no significant difference between the FeSO<sub>4</sub> and Fe-Gly treatments in the direction of BL to AP. Similar results were also found on Fe transport rate of Fe-Gly and FeSO<sub>4</sub> across Caco-2 monolayers. The apparent permeability coefficient of Fe-Gly was significantly higher than that of FeSO<sub>4</sub> in the direction of AP to BL. The ratio of the apparent permeability coefficient between AP to BL and BL to AP was greater than 1.0 for both forms of Fe resources. The Fe transport amount in Fe-Gly treatment was higher than that in FeSO<sub>4</sub> treatment, no matter the incubation temperature was 37°C or 4°C. However, it significantly decreased, when the incubation temperature dropped from 37°C to 4°C. The results indicate that Fe of Fe-Gly can be easily absorbed than FeSO<sub>4</sub> in Caco2 cells. © 2013 Friends Science Publishers

**Keywords:** Fe-Gly; FeSO<sub>4</sub>; Caco-2 Cell; Concentration; Time; Temperature

### Introduction

Iron (Fe) plays an important role in physiological functions, for instance, carries oxygen, forms a part of the oxygen-carrying proteins (hemoglobin and myoglobin) and serves as a cofactor in enzymes that involving in oxidation-reduction reactions (Glahn *et al.*, 2002; Kloots *et al.*, 2004). In terms of oxygen transport and cellular respiration, Fe is an indispensable element for fish (Javed and Saeed, 2010). In order to meet the growth requirement, an addition of 38 to 80 mg of Fe was given to the diet of swine or poultry (Kratzer *et al.*, 1994; 1998). However, studies have shown that a relatively low absorption of nonheme Fe in maize, soybeans and wheat, ranging from 2 to 20% (Hallberg *et al.*, 1997; Tapiero *et al.*, 2001). Therefore, many Fe additives were used in the animal diets to provide enough Fe source, such as Fe sulfate, Fe carbonate, Fe proteinate, and Fe chelate.

The bioavailability values vary greatly among different Fe sources. Compared with ferrous sulfate (FeSO<sub>4</sub>), chelated or proteinated source of Fe had higher bioavailability which was 125 to 185% at the same level of FeSO<sub>4</sub> (Henry *et al.*, 1995). Langini *et al.* (1988) reported that the Fe absorptions of weanling rats which given

infant formula labeled with [<sup>59</sup>Fe] glycine and [<sup>59</sup>Fe] sulfate were 30.9 and 15.8%, respectively. Layrisse *et al.* (2000) found that the absorption rate of Fe in iron glycine chelate (Fe-Gly) was almost two times of that in FeSO<sub>4</sub>. We also reported a better bioavailability in ferrous glycine than ferrous sulfate in piglet (Feng *et al.*, 2007; 2009; Ma *et al.*, 2012). However, the data of absorption characteristics between Fe-Gly and FeSO<sub>4</sub> is still limited.

Caco-2 line that derived from the human colon adenocarcinoma is able to differentiate spontaneously (Sanchez *et al.*, 1996). It can present many absorption characteristics analogous to intestinal cells during culture, such as formatting a monolayer of the cells and expressing several morphological and functional characteristics of the mature enterocyte (Rousset, 1986; Artursson, 1990; Gan *et al.*, 1997). Caco-2 cell model has been used extensively in numerous of biological, biochemical and toxicological studies, as well as in the intake or transport study of trace elements, such as iron, zinc, copper, chromium, and so on (Zödlä *et al.*, 2005; He *et al.*, 2008; Villarroel *et al.*, 2011). Thus, the purpose of the present study was to compare the absorption characteristics between Fe-Gly and FeSO<sub>4</sub> using the Caco-2 cell lines.

## Materials and Methods

### Cells

Caco-2 cells during 20-40 serial passages (Shanghai Institute of Biochemistry and Cell Biology, SIBS, CAS) were cultured according to the method described by Mazariegos *et al.* (2004). Briefly, cells were cultured in 25 cm<sup>3</sup> flasks with Dulbecco's minimum essential medium (DMEM, Gibco) which supplemented with 10% fetal bovine serum (FBS), 4.5 g/L glucose, 2% L-glutamine, 1% non-essential amino acids and 100 U/L penicillin/streptomycin. The incubation condition was 37°C, 5% CO<sub>2</sub> and 90% relative humidity. Suspended the Caco-2 cells, which were in the logarithmic growth phase and reseeded them on the polycarbonate membrane of 6-Transwell plug-in Petri dish at a density of 1×10<sup>4</sup> cells/cm<sup>2</sup>.

### Assessment of Caco-2 Model

The cells were evaluated for study use on the 20-22<sup>th</sup> days. Caco-2 cells were similar to the epithelial cells morphologically (observed by inverted phase contrast microscope, Olympus CKX41, Japan), transmembrane resistance value (466.75 Ω·cm<sup>2</sup>) and mannitol permeation rate (0.85%) could meet the requirements of tightness, integrity and permeability. Besides, cells differentiated well with polarity, which tested by alkaline phosphatase activity. Therefore, Caco-2 model could be used for intestinal absorption *in vitro*.

### Fe Solutions

Fe-Gly and FeSO<sub>4</sub> were made into 1 mol/L stock solution with double-stilled water. Filtrated the stock solution with aseptic Millipore filter and diluted into different multiple with D'Hanks buffer. Atomic absorption spectrometry (Shimadzu AA-6501; Paleologos *et al.*, 2002) was used for the measurement of Fe concentration.

### Transport Assays in Caco-2 Cells

We conducted the bidirectional transport of FeSO<sub>4</sub> and Fe-Gly using caco-2 monolayers prepared as above. Transports from the apical side (AP) to the basolateral side (BL): Sample solutions (1.5 mL) were placed on the apical side of Caco-2 cells, served as the donor chamber, while the basolateral side was supplemented with D'Hanks (2.6 mL) buffer as a receiver. Whereas, transports from BL to AP: BL with 2.6 mL sample served as the donor chamber and AP with 1.5 mL D'Hanks buffer as the receiver. Triplicate wells were used for each treatment.

### Transport Study on Concentration

Sample solutions (1.5 mL) containing different Fe concentration (0.5, 5, 10 and 20 μmol/L) were placed on the side of the donor chamber, while the side of the receiver chamber was supplemented with 2.6 mL D'Hanks buffer.

The experiment was performed in the oscillation sink with 37°C constant temperature and at a speed of 50 rpm. Transport of Fe across the cell monolayer was tested by withdrawing 200 μL of sample from the receiver chamber at different time points, and an equivalent amount of pre-warmed D'Hanks buffer was immediately given to replace this volume. The experiment was ended after 120 min.

### Transport Studies on Time and Temperature

As for the study of transports across Caco-2 monolayers on time and temperature, we collected 200 μL of sample from the receiver chamber at the specific time points (30, 60, 90 and 120 min) and just changed the temperature condition from 37°C to 4°C, respectively. The other details were same as the different Fe concentration study.

### Calculations

Apparent permeability coefficient ( $P_{app}$ , cm/s)

$P_{app}$  was calculated by the following formula:

$$P_{app} = \frac{dQ}{dt \times 60 \times A \times C_0}$$

Where,  $dQ/dt$  (μmol/min) is the flux rate of mass transport across the monolayers,  $A$  is the surface of insert membrane (4.7 cm<sup>2</sup>),  $C_0$  is the initial concentration (μmol/mL) of sample in the donor chamber.

The following equation was applied to revise the values for minimizing sampling errors.

$$TR_{cum} = A_n + \frac{V_{sn}}{V_R} \sum_{i=0}^{n-1} A_i$$

Where,  $TR_{cum}$  is the revised mass transport across the monolayers,  $A_n$  is the transport amount, which is actual measured,  $V_{sn}$  is the volume of collected sample,  $V_R$  is the volume of the donor chamber.

The transport rate is calculated using the following equation:

$$\text{Transport rate(\%)} = \frac{\text{Fe transport amounts in the receiver chamber}}{\text{Fe transport amounts in the donor compartment}} \times 100\%$$

### Statistical Analysis

Values presented in the study are given as the mean ± SEM. One-way analysis of variance (ANOVA) and paired comparison of LSD were used in analyzing the effects between different treatments in SPSS 11.5 software. P value of < 0.05 was considered significant, while < 0.01 were regarded extremely significant.

## Results

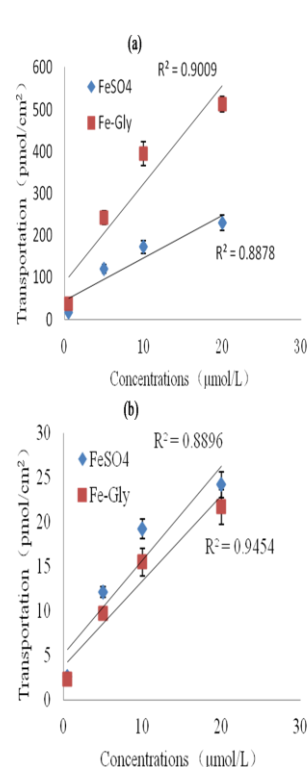
### Effect of Concentration on Fe Transport

The bidirectional transports of Fe-Gly and FeSO<sub>4</sub> are concentration-dependent at the temperature of 37°C (Fig. 1).

**Table 1:** Apparent permeability coefficient ( $P_{app}$ ) of Fe-Gly and FeSO<sub>4</sub> at different concentrations across Caco-2 cell monolayers

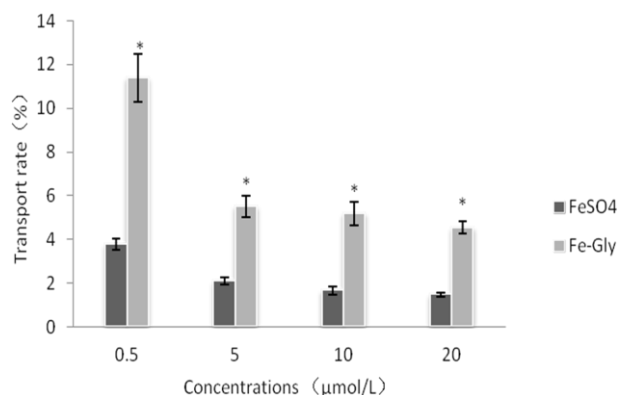
Concentration ( $\mu\text{mol/L}$ )	Apparent permeability coefficient ( $P_{app}$ , $\times 10^{-6}\text{cm/s}$ )					
	FeSO <sub>4</sub>			Fe-Gly		
	AP→BL	BL→AP	R*	AP→BL	BL→AP	R*
0.5	4.70±0.67 <sup>b</sup>	0.72±0.05 <sup>β</sup>	6.50	10.40±0.62 <sup>a</sup>	0.64±0.08 <sup>a</sup>	16.14
5	3.37±0.14 <sup>b</sup>	0.34±0.02	9.99	6.74±0.21 <sup>a</sup>	0.33±0.04	20.71
10	2.39±0.12 <sup>b</sup>	0.27±0.0	8.96	5.48±0.25 <sup>a</sup>	0.21±0.03	25.43
20	1.60±0.11 <sup>b</sup>	0.18±0.03	8.77	3.55±0.39 <sup>a</sup>	0.15±0.04	23.57

Note: Experiments were conducted 120 min at 37°C. The ab or aβ represents the differences between Fe-Gly and FeSO<sub>4</sub> treatments under the same concentration and direction, without common letters are considered significantly ( $P < 0.05$ ). R represents the apparent permeability coefficient ratio

**Fig. 1:** Effects of concentration (0.5–20  $\mu\text{mol/L}$ ) on Fe bidirectional transport across Caco-2 monolayers (37°C, 120 min). (a) shows the transport of Fe-Gly and FeSO<sub>4</sub> from apical to basolateral side. (b) is the results in the inverse direction. The regression lines indicate the positive correlation between transport amount and concentration of Fe

Fe transport amount in Fe-Gly treatment is higher than that in FeSO<sub>4</sub> from AP to BL side, but there is no obvious difference in the direction of BL to AP. The same results were also found on Fe transport rate of Fe-Gly and FeSO<sub>4</sub> across Caco-2 monolayers (Fig. 2).

Table 1 shows the  $P_{app}$  of Fe-Gly and FeSO<sub>4</sub> at different concentrations across Caco-2 cell monolayers. The  $P_{app}$  of Fe-Gly is significantly higher than that of FeSO<sub>4</sub> in the direction of AP to BL. Besides, the ratio of the  $p_{app}$  value between AP to BL and BL to AP is greater than 1.0 for the both forms of iron resources.

**Fig. 2:** Transport rate of Fe-Gly and FeSO<sub>4</sub> across Caco-2 monolayers (AP→BL). Bars within an Fe concentration with asterisk are significantly ( $P < 0.05$ ) different

### Effect of Time and Temperature on Fe Transport

Fe transport amount of Fe-Gly and FeSO<sub>4</sub> across Caco-2 monolayers are linear increased with the time prolonged (Fig. 3). The bidirectional Fe transport of Fe-Gly and FeSO<sub>4</sub> are time-dependent at the temperature of 37°C. It also showed similar pattern as the concentration on Fe transportation, Fe transport amount in Fe-Gly is higher than in FeSO<sub>4</sub> from AP to BL side.

Effect of temperature on Fe transport was shown in Table 2. Compared with FeSO<sub>4</sub>, Fe transport amount in Fe-Gly treatments were both higher when incubated in 37°C or 4°C. However, it decreased ( $P < 0.05$ ) when the incubation temperature dropped from 37°C to 4°C.

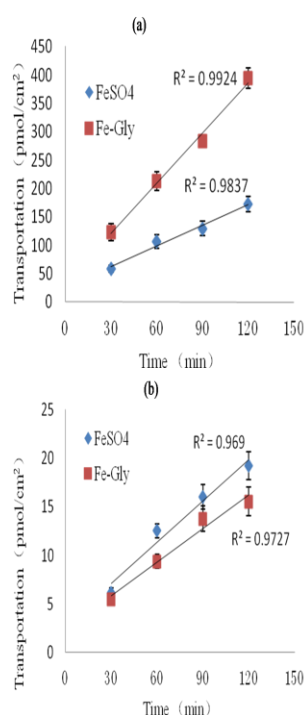
### Discussion

Iron amino acid chelate has been proven to be one of the original models for animals to absorb Fe compound supported by studies on Fe absorption mechanism (Saltman, 1965). Many studies have showed that Fe-Gly has a high bioavailability in the bodies of rats, human beings and other animals, compared with FeSO<sub>4</sub>, (Ashmead, 2001; Allen, 2002). Langini *et al.* (1988) feed weaning rats with infant formula food which added the same level of Fe-Gly or FeSO<sub>4</sub> marked by isotope [<sup>59</sup>Fe]. The study showed that the iron absorption rate of FeSO<sub>4</sub> was 15.8%, while the rate of Fe-Gly was 30.9%. Layrisse *et al.* (2000) reported that the

**Table 2:** Effect of temperature on transport of Fe-Gly and FeSO<sub>4</sub> across Caco-2 monolayers (10 µmol/L, 37°C, 120 min)

Transport amount (pmol/cm <sup>2</sup> )	AP→BL	
	37°C	4°C
FeSO <sub>4</sub>	172.34±5.62 <sup>a</sup>	120.41±2.79 <sup>b</sup>
Fe-Gly	394.55±12.31 <sup>a</sup>	278.55±10.40 <sup>b</sup>

Note: Measurements without common letters in the same row are considered significantly different ( $P<0.05$ )

**Fig. 3:** Effect of time (0-120 min) on Fe transport across Caco-2 cell monolayers (37°C, 10µmol/L, 120 min). (a) shows the transport of Fe-Gly and FeSO<sub>4</sub> from apical to basolateral side. (b) is the results in the inverse direction. The regression lines indicate the positive correlation between transport amount and concentration of Fe

Fe absorption rate of Fe-Gly was higher than that of FeSO<sub>4</sub> by nearly 2-fold in human being body. Ashmead (2001) supplied Fe-Gly and FeSO<sub>4</sub> as Fe fortification for anemia, at the dose of 5 mg/kg weight. After 28 days experiment, the apparent biological utilization rate of Fe-Gly was 90.9%, but the rate of FeSO<sub>4</sub> was only 26.75% with plasma ferritin were monitored as biomarker. Nielsen *et al.* (2005) conducted an experiment treating anemia with Fe-Gly offered as Fe fortificants for nutrition. After 6-week therapy, they found that the mean heme value of patients in the Fe-Gly treatment group was significantly higher than the value of inorganic iron group (12.1±1.8; 10.7±1.7 g/dl). In America, Fe-Gly as one of new iron fortificants for nutrition has been applied to milks of infant and food (Fox *et al.*, 1998; Giorgini *et al.*, 2001).

This study was designed to compare the bidirectional transport of Fe-Gly and FeSO<sub>4</sub> using Caco-2 cells. It proved that Fe of Fe-Gly can be easily absorbed than that of FeSO<sub>4</sub> in Caco-2 cells. We found that the transepithelial transport of Fe-Gly and FeSO<sub>4</sub> were concentration- and time-dependent from both directions. And the transport amounts were much greater in the direction of AP to BL than those in BL to AP transports. The  $P_{app}$  of Fe-Gly and FeSO<sub>4</sub> decreased as the concentration increased and the  $P_{app}$  of transport from AP to BL divide by that in inverse direction is greater than 1.0 for the both two forms of iron sources. The culture temperature has significant effect on transport amount. These results suggest that the absorption of FeSO<sub>4</sub> and Fe-Gly in Caco-2 cells is mainly through active transport. It was also found that the transport amount of Fe-Gly was notably higher than that of FeSO<sub>4</sub> at the same concentration or temperature. All the results taken into account, we speculate that Fe-Gly may have specific or non-specific intestinal active transit system, except for some ionic iron dissociating from Fe-Gly before uptake and being transported through the apical membrane follow the same pattern as FeSO<sub>4</sub> absorption.

The absorption mechanism of Fe-Gly is still unknown exactly. Zhu *et al.* (2006) proposed at least some iron dissociates from EDTA and is reduced just as simple inorganic iron at the brush border membrane of the enterocyte when study iron uptake by Caco-2 cells from NaFe-EDTA and FeSO<sub>4</sub>. Pizarro *et al.* (2002) found that Fe-Gly competes for the absorption pathway of nonheme Fe. They enter the common nonheme Fe pool and activate the same transporter on the cells for transportation. They concluded that one reason for better absorption of Fe-Gly is glycine can chelate and protect Fe from inhibitors.

In conclusion, our work confirmed that part of Fe-Gly absorbed by the same pattern as FeSO<sub>4</sub> absorption mechanism, but it is likely that Fe-Gly is also using specific or non-specific intestinal active transit system. For further study, transport kinetics of Fe-Gly and relevant proteins of Fe transporter should be tested for revealing the mechanism.

## Acknowledgements

This work was supported by the Project supported by the National Natural Science foundation (No.31272398), New-Century Training Program Foundation for Talents from the Ministry of Education of China (No. NCET-10-0727), and the Natural Science Foundation for Distinguished Young Scholars of Zhejiang province, China (No. R3110085).

## References

- Artursson, P., 1990. Epithelial transport of drugs in cell culture I: A model for studying the passive diffusion of drugs over intestinal absorptive (Caco-2) cells. *J. Pharm. Sci.*, 79: 476-482
- Ashmead, S.D., 2001. The chemistry of ferrous bis-glycinate chelate. *Arch. Latinoam. Nutr.*, 51: 7-12

- Allen, L.H., 2002. Advantages and limitations of iron amino acid chelates as iron fortificants. *Nutr. Rev.*, 60: S18–S21
- Fox, T.E., J. Eagles and S.J. Faiweather-Tait, 1998. Bioavailability of iron glycine as a fortificant in infant foods. *Amer. J. Clin.*, 67: 664–668
- Feng, J., W.Q. Ma, Z.R. Xu, Y.Z. Wang and J.X. Liu, 2007. Effects iron glycine chelate on growth, haematological and immunological characteristics in weanling pigs. *Anim. Feed Sci. Tech.*, 134:261–272
- Feng, J., W.Q. Ma, Z.R. Xu, J.X. He, Y.Z. Wang and J.X. Liu, 2009. The effect of iron glycine chelate on tissue mineral levels, fecal mineral concentration, and liver antioxidant enzyme activity in weanling pigs. *Anim Feed Sci. Technol.*, 150: 106–113
- Gan, L.S.L. and D.R. Thakker, 1997. Applications of the Caco-2 model in the design and development of orally active drugs: elucidation of biochemical and physical barriers posed by the intestinal epithelium. *Adv. Drug. Deli. Rev.*, 23: 77–98
- Giorgini, E., M. Fisberg, R.A. Paula, A.M. Ferreira, J. Valle and J.A. Braga, 2001. The use of sweet rolls fortified with iron-bis-glycinate chelate in the prevention of iron deficiency anemia in preschool children. *Arch. Lothinoam Nutr.*, 51: 48–53
- Glahn, R.P., Z.Q. Cheng and M.R. Welch, 2002. Comparison of iron bioavailability from 15 rice genotypes: studies using an *in vitro* digestion/Caco-2 cell culture model. *J. Agric. Food Chem.*, 50: 3586–3591
- Henry, P.R. and E.R. Miller, 1995. Iron availability. In: *Bioavail. Nutation Animal*, pp: 169–199. Ammerman, C.B., D.H. Baker and A.S. Lewis (eds.). Academic Press, San Diego, California, USA
- Hallberg, L., L. Hulten and E. Gramatkovski, 1997. Iron absorption from the whole diet in men: how effective is the regulation of iron absorption? *Amer. J. Clin. Nutr.*, 66: 347–356
- He, W., Y. Feng, X. Li, Y. Wei and X. Yang, 2008. Availability and toxicity of Fe(II) and Fe(III) in Caco-2 cells. *J. Zhejiang Univ. Sci. B.*, 9: 707–712
- Javed, M. and M.A. Saeed, 2010. Growth and Bioaccumulation of Iron in the Body Organs of Catla catla, Labeo rohita and Cirrhina mrigala during Chronic Exposures. *Int. J. Agric. Biol.*, 6: 881–886
- Kratzer, F.H., J.D. Latshaw, S.L. Leeson, E.T. Moran Jr., C.M. Parsons, J.L. Sell and P.W. Waldroup, 1994. *Nutrient Requirements of Poultry*, 9<sup>th</sup> edition. National Academy Press, Washington, DC, USA
- Kratzer, F.H., J.D. Latshaw, S.L. Leeson, E.T. Moran Jr., C.M. Parsons, J.L. Sell and P.W. Waldroup, 1998. *Nutrient Requirements of Swine*, 10<sup>th</sup> edition. National Academy Press, Washington, DC, USA
- Kloots, W., D.O. den Kamp and L. Abrahamse, 2004. *In vitro* iron availability from iron-fortified whole-grain wheat flour. *J. Agric. Food Chem.*, 52: 8132–8136
- Langini, S., N. Carbone, M. Galdi, M.E. Barrio rendo, M.L. Portela, R. Caro and M.E. Valencía, 1988. Ferric glycinate iron bioavailability for rats, as determined by extrinsic radioisotopic labeling of infant formulas. *Nutr. Rep. Int.*, 38: 729–735
- Layrisse, M., M.N. García-Casal, L. Solano, M.A. Baron, F. Arguello, D. Llovera, J. Ramirez, I. Leets and E. Tropper, 2000. Iron bioavailability in humans from breakfasts enriched with iron bis-glycine chelate, phytates and polyphenols. *J. Nutr.*, 130: 2195–2199
- Mazariegos, D.I., F. Pizarro, M. Olivares, M. Olivares, M.T. Nunez and M. Arredondo, 2004. The mechanisms for regulating absorption of Fe bis-glycine chelate and Fe-ascorbate in Caco-2 cells are similar. *J. Nutr.*, 134: 395–398
- Ma, W.Q., H. Sun, Y. Zhou, J. Wu and J. Feng, 2012. Effects of iron glycine chelate on growth, tissue mineral concentrations, fecal mineral excretion, and liver antioxidant enzyme activities in broilers. *Biol. Trace Elem. Res.*, 149: 204–211
- Nielsen, P., R. Kongi and P. Buggisch, 2005. Bioavailability of oral iron drugs as judged by a <sup>59</sup>Fe-whole-body counting technique in patients with iron deficiency anaemia. Therapeutic efficacy of iron (II)-glycine sulfate. *Arzneimittel-Forsch.*, 55: 376–381
- Paleologos, E.K., D.L. Giokas, S.M. Tzouwaras-Karayanni and M.I. Karayannis, 2002. Micelle mediated methodology for the determination of free and bound iron in wines by flame atomic absorption spectrometry. *Anal. Chim. Acta*, 458: 241–248.
- Pizarro, F., M. Olivares, E. Hertrampf, D. Mazariegos, M. Arredondo, A. Letelier and V. Gidi, 2002. Iron bis-glycine chelate competes for the nonheme-iron absorption Pathway. *Amer. J. Clin. Nutr.*, 76: 577–581
- Rousset, M., 1986. The human colon carcinoma cell lines HT-29 and Caco-2: Two *in vitro* models for the study of intestinal cell differentiation. *Biochimie*, 68: 1035–1040
- Saltman P., 1965. The role of chelation in iron metabolism. *J. Chem. Edu.*, 42: 682
- Sanchez, L., M. Ismail, F.Y. Liew and J.H. Brock, 1996. Iron transport across Caco-2 cell monolayers. Effect of transferrin, lactoferrin and nitric oxide. *Biochim. Biophys. Acta*, 1289: 291–297
- Tapiero, H., L. Gate and K.D. Tew, 2001. Iron: deficiencies and requirements. *Biomed. Pharmacother*, 55: 324–332
- Villarreal, P., S. Flores, F. Pizarro, D.L. de Romaña and M. Arredondo, 2011. Effect of dietary protein on heme iron uptake by Caco-2 cells. *Eur. J. Nutr.*, 50: 637–643
- Zödl, B., M. Zeinerb, P. Paukovitsa, I. Steffanb, W. Marktla and C. Ekmekcioglua, 2005. Iron uptake and toxicity in Caco-2 cells. *Microchem. J.*, 79: 393–397
- Zhu, L., R.P. Glahn, C.K. Yeung and D.D. Miller, 2006. Iron Uptake by Caco-2 Cells from NaFeEDTA and FeSO<sub>4</sub>: Effects of Ascorbic Acid, pH, and a Fe(II) Chelating Agent. *J. Agric. Food Chem.*, 54: 7924–7928

(Received 31 August 2012; Accepted 17 October 2012)