



Full Length Article

## Age-Stage, Two-Sex Life Table Study of the Effects of Sub-Lethal Concentrations of Novaluron on *Earias vittella* (Lepidoptera: Noctuidae)

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### Abstract

*Earias vittella* (F) is an important insect pest of cotton (*Gossypium hirsutum*) and okra (*Abelmoschus esculentus*) in Pakistan. The current study was carried out to explore the effects of sub-lethal concentrations of novaluron on the life table parameters of the pest. Bioassays were performed to assess the sub-lethal concentrations (LC<sub>20</sub> and LC<sub>50</sub>) of the novaluron and its effects on the demographic parameters of the *E. vittella*. Age-stage, two-sex life table theory was applied to interpret the data for population parameters of *E. vittella*. In the current study, the LC<sub>20</sub> and LC<sub>50</sub> were calculated as 2.224 ppm and 9.837 ppm, respectively. The results showed that in novaluron treated samples rates of all biological parameters decreased whereas the larval, pupal period and mean generation time were increased. The intrinsic rate of increase remained high in control as 0.166 d<sup>-1</sup> in comparison with LC<sub>50</sub> as 0.128 d<sup>-1</sup>. The net reproductive rate ranged from 94.542 offsprings per individual (control) to 61.228 offsprings per individual (LC<sub>50</sub>). Fecundity was dropped in insects treated with sub-lethal concentrations from 330.9 eggs per female (control) to 238.11 eggs per female (LC<sub>50</sub>). This study revealed that the sub-lethal concentrations of novaluron significantly decreased the biological rate of *E. vittella* under laboratory conditions and suggests that such doses should be practiced in the fields for proper integrated pest management strategies. © 2022 Friends Science Publishers

**Keywords:** *Eairas vittella*; Sub-lethal concentration; Novaluron; Insect growth regulator; Two-sex life table

### Introduction

*Eairas vittella* (F.) (Noctuidae: Lepidoptera), also known as spotted bollworm, is a notorious polyphagous pest of many malvaceous crops (Aziz *et al.* 2012). Some of its major host crops include *Gossypium* spp. (cotton), *Abelmoschus esculentus* (okra), *Abutilon indicum*, *Hibiscus cannabinus*, *Althaearosea* (hollyhock) and *Malwa parviflora* (sonchal) (Rasool *et al.* 2002; Jan *et al.* 2015; Rahman *et al.* 2016). The spotted bollworm is active throughout the year and have 6 to 8 generations during each year. Several buds and bolls are damaged by a single larva in its life span. In cotton, it pupates in bolls and reduces the boll growth (Aziz *et al.* 2011; Jan *et al.* 2015) commonly the buds, flowers, and fruits are attacked by second instar larvae and results in the reduction of quality and quantity. It may reduce the yield up to 50% in cotton and about 69% in okra (Aziz *et al.* 2011).

The use of chemicals plays a key role in the management of pests in fields, resulting in a low risk of yield loss (Popp and Hantos 2011). *E. vittella* infestation has been managed using different insecticides including, pyrethroids and organophosphates (Praveen *et al.* 2007; Umrao *et al.*

2013). Irregular and massive application of such chemicals has resulted in resistance development in field pests (Abbas *et al.* 2014; Gulzar and Wright 2014; Abbas and Shad 2015; Ahmad *et al.* 2019). Novaluron is one of the recent insect growth regulators (IGR) that belong to the insecticidal group, benzoylphenyl urea. It is a chitin synthesis inhibitor that acts through ingestion and contact. It targets the larval stage of insects which synthesize chitin actively (Lohmeyer and Pound 2012). The residual activity of novaluron in field conditions depends upon the environmental conditions and ranges from 10 to 30 days (Ishaaya *et al.* 2003). It has a very low toxic effect on mammals, birds, earthworms (Ishaaya *et al.* 2007) and adults of non-targeted beneficial insect species (Cutler *et al.* 2005).

The application of pesticides in fields does not kill all the pest populations with immediate effects, so over time the pesticide decreases and, as a result, the sublethal effects including behavioral and physiological changes in pests can occur (Rehan and Freed 2015). These sublethal concentrations of pesticides can significantly alter the adult development, adult insect weight, larval and pupa periods and reproduction parameters of the insect (Han *et al.* 2012).

For a comprehensive pesticide evaluation, only acute toxicity is not enough, but the sublethal effects may also be included (Zhang *et al.* 2015). Previously, Rahmani and Bandani (2013) found that the sub-lethal doses of thiamethoxam chemical significantly altered the different *Hippodamia variegata* population parameters in adverse modus. This study was planned to elucidate the sublethal effects of the novaluron on population parameters of *E. vittella* by application of Age-stage, two-sex life table theory. The finding of this study may be helpful to monitor the insecticide resistance in *E. vittella* to insect growth regulators (IGRs) in fields and develop improved integrated pest management strategies.

## Materials and Methods

### Insect culture and insecticides

Laboratory culture of spotted bollworm (*E. vittella*) was established from the larvae collected from okra fields in the surrounding of Rawalpindi, Taxila and Attock. The infested pods from the fields were transferred into transparent plastic jars (20 cm length and 10 cm wide) and were kept under control conditions ( $27 \pm 2^\circ\text{C}$ ,  $60 \pm 5\%$  R.H. photoperiod of 16 L: 8 D). Insect culture was maintained on okra fruits as described by Al-Mehmmady (2000). Fresh okra fruits were washed thoroughly by tap water and air-dried before feeding to the neonates. The okra fruits were cut into 0.5–1.0 cm pieces, 4–5 larvae were released per piece and placed in the plastic container. The food was replaced daily till pupation. The larvae were carefully removed from the okra pods and the excreta were cleaned. The pupae were shifted to another plastic jar (10 cm length and 5 cm width) until adult emergence. The emerged adults were shifted to adult cages and fed with 10% sugar solution. Nappy strips were hanged in the adult cages as oviposition sites, these strips were replaced regularly when eggs were observed.

Novaluron, insect growth regulator, (Corvus®) FMC Pvt. Ltd. was used at the recommended dose along with different concentrations against the 1<sup>st</sup> instar larvae of spotted bollworm (*E. vittella*) to investigate the larvicidal effects.

### Bioassays

Bioassays were performed by using diet emersion method and toxicity of novaluron was checked against 1<sup>st</sup> larval instar of spotted bollworm (*E. vitella*). Different concentrations of novaluron by serial dilutions ( $\text{mg L}^{-1}$ ) of stock solution were prepared using distilled water. Fresh okra fruits were dipped separately in each concentration for 10 sec and then dried for 10 min at room temperature. Five okra fruits were used in one replication. Two 1<sup>st</sup> instar larvae were released on each okra pods in all replications treated with different concentrations. Each treatment was replicated four times. Distilled water was used as a control treatment. All the treated larvae were kept under controlled

conditions (Temperature of  $25 \pm 2^\circ\text{C}$ ; R. H. of  $65 \pm 5\%$ ). Mortality data were assessed after 72 h.

### Sublethal effects on demographic parameters of *E. vittella*

In life table study, 210 eggs were used, which were collected after 24 h of deposition by females of the laboratory population. Three treatments (control, LC<sub>20</sub>, and LC<sub>50</sub>) were prepared for this experiment. Seventy eggs were treated with each treatment. Each egg is an individual petri dish that was considered as one replicate (Huang and Chi 2013; Zhang *et al.* 2015). All the Petri dishes were kept under controlled conditions. The egg hatching data were recorded daily. The neonates from control, LC<sub>20</sub> and LC<sub>50</sub> were shifted on the okra pods treated with control, LC<sub>20</sub> and LC<sub>50</sub>, respectively. The larval development was observed daily and fresh okra pods were provided after each instar. Pupae were removed and placed in new Petri dishes until emergence. After the adult emergence, they were paired (one male and one female) and transferred to individual plastic containers for oviposition. The adults were checked daily for oviposition and transferred to new containers for egg-laying. The fecundity and survival rate of the adults have assessed until the death of the adults.

### Data analysis

The LC values were calculated based on mortality data by using R Statistical Software version 2.9.0 (R Development Core Team 2009). The data regarding different stage development periods, survival rate and fecundity along with oviposition periods were analyzed using Age-stage, two-sex life table theory (Chi and Liu 1985; Chi 1988) with TWO SEX-MS Chart software (Chi 2017). Means of the biological parameters were compared by using 100,000 bootstrap techniques to achieve stable SE estimates (Huang and Chi 2013). The curves for age-specific survival rate, fecundity, life expectancy and reproductive values were generated by using Sigma Plot 14.0. The net reproductive rate was calculated as:

$$R_0 = \sum_{x=0}^{\infty} l_x m_x$$

The intrinsic rate of increase ( $r$ ) is calculated by using the iterative bisection method from:

$$\sum_{x=0}^{\infty} e^{-r(x+1)} l_x m_x = 1$$

With age indexed from zero (Goodman 1982). The mean generation time ( $T$ ) is calculated as follow:

$$T = \frac{\ln R_0}{r}$$

The Gross reproductive rate (GRR) is calculated by the

formula as follow:

$$GRR = \sum_{x=0}^{\infty} m_x$$

The age-specific survival rate ( $l_x$ ) and age-specific fecundity ( $m_x$ ) were given as:

$$l_x = \sum_{j=1}^k S_{xj}$$

$$m_x = \frac{\sum_{j=1}^k S_{xj} f_{xj}}{\sum_{j=1}^k S_{xj}}$$

## Results

### Toxicity bioassays

The toxicity of novaluron against the 1<sup>st</sup> instar of *E. vittella* after 72 h is given in Table 1. The LC<sub>20</sub> and LC<sub>50</sub> were calculated as 2.224 mg L<sup>-1</sup> and 9.837 mg L<sup>-1</sup>, respectively.

### Sub-lethal effects of novaluron on biological parameters of *E. vittella*

The developmental periods, fecundity and adult longevity of both males and females of *E. vittella* treated with sub-lethal concentrations (LC<sub>20</sub> and LC<sub>50</sub>) of novaluron are given in Table 2. Egg duration was significantly ( $P \leq 0.005$ ,  $df=2$ ,  $F=87.61$ ) prolonged when treated with sub-lethal concentrations (4.26 days and 4.07 days for LC<sub>50</sub> and LC<sub>20</sub> respectively of novaluron as compared with the control (3.66 days). Total larval time was also increased by treating with sub-lethal concentrations (LC<sub>50</sub> and LC<sub>20</sub>) as 12.54 days and 11.23 days respectively ( $P \leq 0.005$ ,  $df=2$ ,  $F=117.24$ ) as compared with control (11.03 days). No significant variation was noted between LC<sub>20</sub> (11.35 days) and LC<sub>50</sub> (11.47 days) in terms of the pupal period but differed significantly ( $P \leq 0.005$ ,  $df=2$ ,  $F=122.82$ ) when compared with the untreated larvae (Table 2). Adult male longevity was not significantly different ( $p=0.104$ ,  $df=2$ ,  $F=11.82$ ), while female longevity was significantly ( $P \leq 0.005$ ,  $df=2$ ,  $F=92.87$ ) different ranging from highest (12.7 days) on control to the lowest (7.62 days) on LC<sub>50</sub>. The differences in total pre-oviposition period (TPOP) between sub-lethal concentrations were statistically non-significant ( $P \leq 0.01$ ,  $df=2$ ,  $F=7.45$ ), while a significant difference was found between the control and treated larvae. The number of eggs per female varied significantly ( $P \leq 0.005$ ,  $df=2$ ,  $F=648.22$ ) between sub-lethal concentrations (238.11 and 268.5 eggs per female in LC<sub>50</sub> and LC<sub>20</sub> respectively). The highest fecundity (330.9 eggs per female) was noted for untreated larvae.

### Sub-lethal effects of novaluron on population parameters of *E. vittella*

Novaluron significantly altered the population parameters of

the *E. vittella* (Table 3). To estimate the population parameters, the bootstrap method with 100,000 replicate sample method was used. The intrinsic rate of increase was decreased by treating with both the concentrations of novaluron (0.128 and 0.140 d<sup>-1</sup> for LC<sub>50</sub> and LC<sub>20</sub>, respectively) as compared with the untreated larvae (0.166 d<sup>-1</sup>). A similar trend was found in the finite rate of increase ( $\lambda$ ), as the highest value for  $\lambda$  was found in control larvae (1.181 d<sup>-1</sup>) which gradually decreases with an increase in concentration from LC<sub>20</sub> to LC<sub>50</sub> as 1.150 and 1.137 d<sup>-1</sup> respectively. A significant decrease was also observed in net reproductive rate after novaluron treatment from being highest on control larvae (94.542 offsprings per individual) to 76.714 and 61.228 offsprings per individual for LC<sub>20</sub> and LC<sub>50</sub> treated larvae respectively. Moreover, the gross reproductive rate (GRR) of LC<sub>20</sub> treated larvae (146.45, offsprings per individual) was significantly similar to that of LC<sub>50</sub> and control. The highest GRR was recorded for control larvae (183.42 offsprings per individual), while the lowest recorded for LC<sub>50</sub> treated larvae (136.77 offsprings per individual) which were both statistically significant to each other. The mean generation time was prolonged in the treated larvae compared to the control larvae. Minimum mean generation time was taken by the control larvae (27.255 d), followed by LC<sub>20</sub> treated larvae (30.967 d). The maximum days were recorded on LC<sub>50</sub> treated larvae (31.917 d). The curves of developmental rates of the individuals showed an overlapped nature showing the differences in their development rates (Fig. 1). Females were emerged late in the population than males, while their survival was longer than males. The LC<sub>20</sub> and LC<sub>50</sub> treatment were observed with less number of larvae as compared with untreated. The longest time for development was noted for LC<sub>50</sub> and LC<sub>20</sub> treated larvae when compared with untreated larvae (Fig. 1).

Age-specific survival rates ( $l_x$ ), age-specific fecundity ( $f_x$ , female), age-specific fecundity for total population and age-specific maternity ( $l_x m_x$ ) are presented in Fig. 2. A significant decline in the curve of  $l_x$  was noted in LC<sub>20</sub> and LC<sub>50</sub> treated larvae after 32 days of the treatment. The untreated group has the highest top peaks of  $f_x$  and  $m_x$  than compared with the LC<sub>20</sub> and LC<sub>50</sub> treated group. A significant variation was shown in the life expectancy ( $e_{xj}$ ) among the treated (LC<sub>20</sub> and LC<sub>50</sub>) and untreated larvae (Fig. 3). The maximum life expectancy of new eggs was recorded in LC<sub>50</sub> (39.0 days), followed by LC<sub>20</sub> (38.0 days) while the minimum life expectancy of eggs was recorded in the untreated group (36.0 days). Reproductive rate ( $v_{xj}$ ) is defined as the measure of dedication to newly coming offspring in the future from age  $x$  to stage  $j$  (Fig. 4). The contribution of males in the population to the next generation was not well defined, therefore the curve for males was not included. A decline was observed in the reproductive values when larvae treated with LC<sub>50</sub> and LC<sub>20</sub> as compared to the untreated larvae. Maximum  $v_{xj}$  was recorded on the untreated group, while the minimum was

**Table 1:** Acute toxicity of novaluron on the 1<sup>st</sup> instar of *E. vittella* after 72 h of treatment

Chemical	n	Concentration mg liter <sup>-1</sup> (95% CL)			Slope ± SE	χ <sup>2</sup> (df)
		LC <sub>10</sub>	LC <sub>20</sub>	LC <sub>50</sub>		
Novaluron	240	1.022 (0.534-2.156)	2.224 (1.162-4.256)	9.837 (5.140-18.827)	1.304 ± 0.144	0.997 (5)

LC (lethal concentration), n (number of samples), CL (confidence level), SE (standard error), χ<sup>2</sup> (Chi square), df (degree of freedom)

**Table 2:** Life table parameters of *E. vittella* treated with sub-lethal concentrations of novaluron

Treatment	Control		LC <sub>20</sub>		LC <sub>50</sub>	
	n	Mean ± SE	n	Mean ± SE	n	Mean ± SE
Egg (days)	70	3.66 ± 0.1 <sup>b</sup>	70	4.07 ± 0.094 <sup>a</sup>	70	4.26 ± 0.125 <sup>a</sup>
1 <sup>st</sup> instar (days)	64	2.28 ± 0.08 <sup>c</sup>	60	2.33 ± 0.088 <sup>b</sup>	58	2.52 ± 0.094 <sup>a</sup>
2 <sup>nd</sup> instar (days)	54	2.19 ± 0.076 <sup>b</sup>	50	2.20 ± 0.082 <sup>b</sup>	56	2.54 ± 0.096 <sup>a</sup>
3 <sup>rd</sup> instar (days)	48	2.17 ± 0.078 <sup>b</sup>	44	2.36 ± 0.105 <sup>a</sup>	46	2.48 ± 0.106 <sup>a</sup>
4 <sup>th</sup> instar (days)	46	2.13 ± 0.072 <sup>b</sup>	42	2.19 ± 0.088 <sup>b</sup>	40	2.53 ± 0.125 <sup>a</sup>
5 <sup>th</sup> instar (days)	46	2.26 ± 0.094 <sup>b</sup>	40	2.15 ± 0.082 <sup>b</sup>	38	2.47 ± 0.125 <sup>a</sup>
Larva (days)	46	11.03 ± 0.028 <sup>c</sup>	40	11.23 ± 0.041 <sup>b</sup>	38	12.54 ± 0.013 <sup>a</sup>
Pupa(days)	41	8.74 ± 0.201 <sup>b</sup>	34	11.35 ± 0.15 <sup>a</sup>	31	11.47 ± 0.151 <sup>a</sup>
Male longevity(days)	18	8.52 ± 2.066 <sup>a</sup>	19	7.72 ± 1.56 <sup>a</sup>	16	7.12 ± 0.331 <sup>a</sup>
Female longevity(days)	23	12.7 ± 2.13 <sup>a</sup>	15	9.04 ± 1.21 <sup>b</sup>	14	7.62 ± 0.242 <sup>c</sup>
APOP	23	1.2 ± 0.133 <sup>a</sup>	15	1.00 ± 0.00 <sup>a</sup>	14	1.00 ± 0.00 <sup>a</sup>
TPOP	23	24.5 ± 0.619 <sup>b</sup>	15	28.3 ± 0.26 <sup>a</sup>	14	29.22 ± 0.464 <sup>a</sup>
Fecundity (eggs/f)	23	330.9 ± 26.41 <sup>a</sup>	15	268.5 ± 19.67 <sup>b</sup>	14	238.11 ± 14.24 <sup>c</sup>

LC (lethal concentration), SE (standard error), n (number of insects exposed). Adult pre oviposition period (APOP), total pre oviposition period (TPOP), means sharing similar letters in a row are not different statistical at 5% probability

**Table 3:** Effect of sub-lethal concentration of novaluron on the biological parameters of *E. vittella*

Population parameters	Control		LC <sub>20</sub>		LC <sub>50</sub>	
	Mean	SE	Mean	SE	Mean	SE
Intrinsic rate of increase ( <i>r</i> )	0.166 <sup>a</sup>	0.011	0.140 <sup>b</sup>	0.009	0.128 <sup>c</sup>	0.010
Finite rate of increase ( $\lambda$ )	1.181 <sup>a</sup>	0.013	1.150 <sup>b</sup>	0.010	1.137 <sup>b</sup>	0.011
Net reproductive rate ( <i>R<sub>n</sub></i> )	94.542 <sup>a</sup>	25.23	76.714 <sup>ab</sup>	20.679	61.228 <sup>b</sup>	17.610
Mean generation time ( <i>T</i> )	27.255 <sup>c</sup>	0.573	30.967 <sup>b</sup>	0.272	31.917 <sup>a</sup>	0.301
Gross reproductive rate ( <i>GRR</i> )	183.42 <sup>a</sup>	8.055	146.45 <sup>ab</sup>	32.83	136.77 <sup>b</sup>	30.553

LC (lethal concentration), SE (standard error), df (degree of freedom), means sharing similar letters in a row are not different statistical at 5% probability

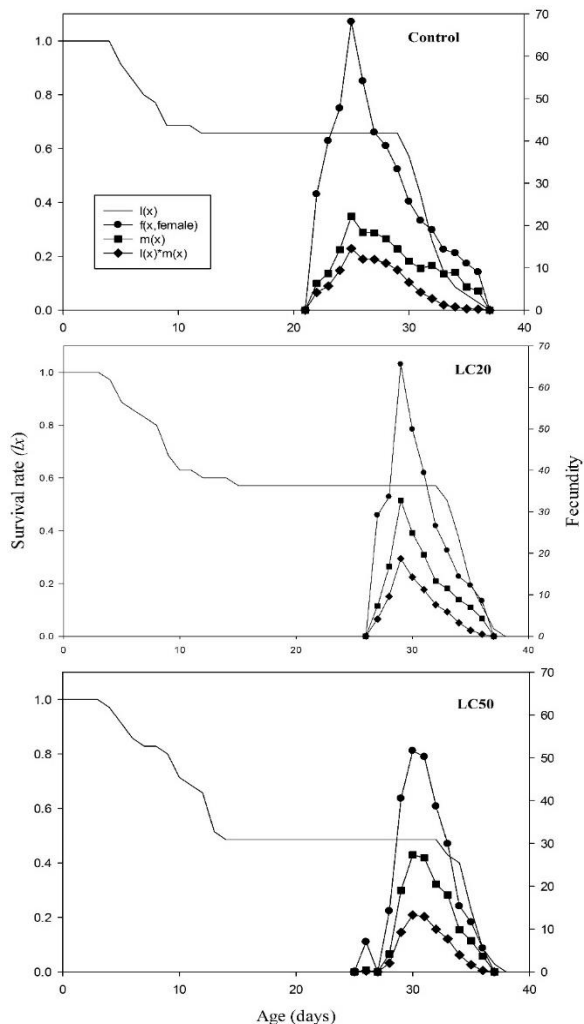
observed in LC<sub>50</sub> treated group.

## Discussion

In the current study, Age-stage, two-sex life table theory was utilized to calculate the population parameters of *E. vittella* exposed to the sublethal concentrations of novaluron. Age-stage, two-sex life table study is a promising way to estimate the population parameters of the pest by considering both the sexes of the existing population (Rahmani and Bandani 2013; Huang and Chi 2013). The intrinsic rate of increase, reproductive rate and total oviposition period are the most important characteristics of the life table study to predict the insect population effected by insecticides (Papachristos and Milonas 2008).

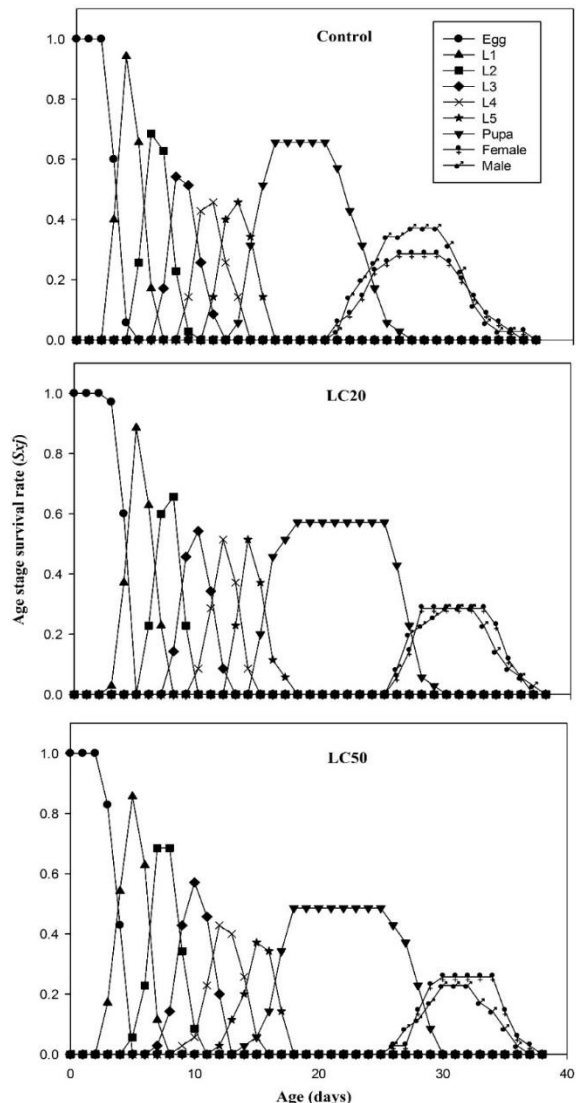
The sub-lethal concentrations (LC<sub>20</sub> and LC<sub>50</sub>) of novaluron for *E. vittella* were calculated as 2.224 and 9.837 mg L<sup>-1</sup>, respectively in this study. Cutler et al. (2005) studied the acute toxicity of novaluron on the second instar of *L. decemlineata* and calculated LC<sub>50</sub> as 18.7 ppm and categorized it as broad-spectrum insecticides. Population parameters of *E. vittella* studied in this paper showed that the sub-lethal (LC<sub>20</sub> and LC<sub>50</sub>) concentrations of novaluron have decreased the intrinsic rate of increase, finite rate of

increase, reproductive value, survival rate, and net reproductive rate, however, the larval period, pupal period and TPOP were increased by the sub-lethal concentrations. The above results proved that the population growth of *E. vittella* was significantly reduced by the sub-lethal concentrations of novaluron. These results are according to the sub-lethal effects of thiamethoxam on *Bradysia odoriphaga* (Zhang et al. 2015). Significantly no difference was found in the adult pre-oviposition period (APOP) among treatments, while a significant variation was found in the total pre-oviposition period (TPOP), which was positively correlated with the intrinsic rate of increase of the *E. vittella*. Sub-lethal concentrations significantly prolonged the larval duration, this may have occurred because of agitations in nerve tissue development by neurotoxic chemical contact (Desneux et al. 2007). Similar results were found when the population of *E. vittella* treated with sublethal doses of lufenuron (Hafeez et al. 2019). The reason for the prolonged larval duration would be that the treated larvae were more intense with the detoxification of the sub-lethal effect of novaluron causing the increased larval period as compared to the control (Meng et al. 2018). The number of eggs per female also reduced to a significant level by the application of sub-lethal concentrations of



**Fig. 2:** Survival rate ( $l_x$ ) and fecundity of *E. vittella* exposed to sublethal concentration of novaluron

novaluron. This showed that the chemical has produced some effects on the ovaries of the females in treated groups (Seth *et al.* 2004; Qu *et al.* 2017). It has been found that exposure of insects to insecticides results in the reduction of ovarioles size, basal oocytes, and firmness of follicular epithelium of armyworm, which was the main reason for reduced fecundity (Perveen and Miyata 2000). The reduction may also be due to changes in the behavioral and physiological effects of the insecticide. The short life span of adults will result in the short mating periods, which will ultimately result in fewer eggs in the field and thus a decline will be found in the field population over time. In this study, the population parameters *viz.*, intrinsic rate of increase, gross reproductive rate, net reproductive rate and finite rate of increase were decreased with the LC<sub>20</sub> and LC<sub>50</sub> treatments. These results are in line with the result of previous studies, in which the intrinsic rate of increase, finite rate of increase and net reproductive rate statistically declined in diamondback moth and cabbage aphid by the

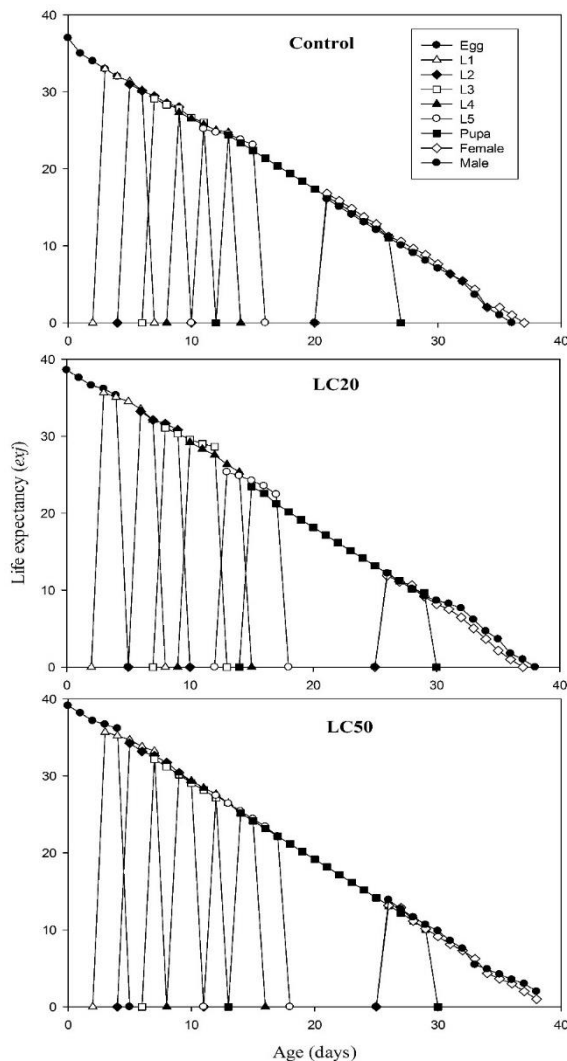


**Fig. 1:** Age-stage specific survival rate ( $s_{xj}$ ) of *E. vittella* exposed to sublethal concentration of novaluron

treatment of spinosad and imidacloprid respectively (Lashkari *et al.* 2007; Yin *et al.* 2008). This study provides a clear knowledge about the population dynamics of *E. vittella* subjected to the sub-lethal effects of the insecticides. These sublethal concentrations of novaluron can be utilized in the field to delay the development rate and reduce the reproduction ability of the females in the pest population. These concentrations will reduce the number of insecticides used in fields, reduce the risk of resistance development in the insect pests and reduce environmental pollution.

**Conclusion**

This study publicized that the novaluron with sub-lethal concentrations significantly decreased the biological rate of *E. vittella* in the laboratory conditions and advocates the



**Fig. 3:** Life expectancy ( $e_{xj}$ ) of *E. vittella* exposed to sublethal concentration of novaluron

implementation of such doses in the fields for incorporation in integrated pest management strategies.

### Author Contributions

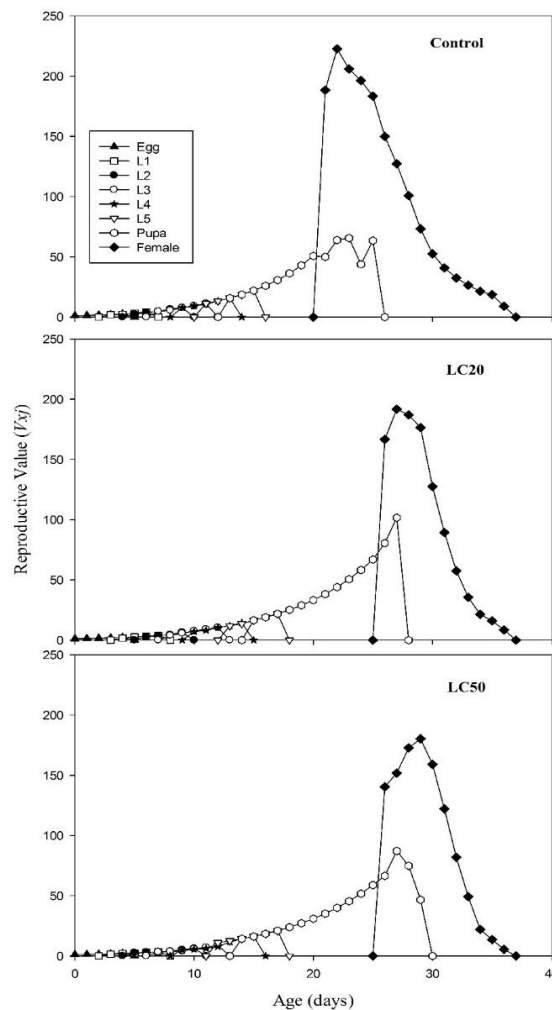
DK, AG, BR and MT designed the research. DK, SK and SA conducted the experiments. BR and DK analyzed the data. DK, BR and IH wrote the manuscript. All the authors read and approved the manuscript.

### Conflict of Interest

Authors declare that there is no conflict of interest

### Funding Source

There is no funding source for the present research



**Fig. 4:** Reproductive value ( $V_{xj}$ ) of *E. vittella* exposed to sublethal concentration of novaluron

### References

- Abbas N, SA Shad (2015). Assessment of resistance risk to lambda-cyhalothrin and cross-resistance to four other insecticides in the house fly, *Musca domestica* L. (Diptera: Muscidae). *Parasitol Res* 114:2629–2637
- Abbas N, SA Shad, M Razaq, A Waheed, M. Aslam (2014). Resistance of *Spodoptera latera* (Lepidoptera: Noctuidae) to profenofos: Relative fitness and cross-resistance. *Crop Prot* 58:49–54
- Ahmad M, B Rasool, M Ahmad, DA Russell (2019). Resistance and synergism of novel insecticides in field populations of cotton bollworm *Helicoverpa armigera* (Lepidoptera: Noctuidae) in Pakistan. *J Econ Entomol* 112:859–871
- Al-Mehmmady RM (2000). Biological studies on the okra moth, *Earias vittella* (F.) (Lepidoptera: Noctuidae) in Jeddah, Saudi Arabia. *Res Bull* 96:5–18
- Aziz MA, MU Hasan, A Ali, J Iqbal (2012). Comparative efficacy of different strategies for management of spotted bollworms, *Earias* spp. on Okra, *Abelmoschus esculentus* (L.). Moench. *Pak J Zool* 44:1203–1208
- Aziz MA, M Hasan A Ali (2011). Impact of abiotic factors on incidence of fruit and shoot infestation of spotted bollworms *Earias* spp. on okra (*Abelmoschus esculentus* L.). *Pak J Zool* 43:863–868

- Chi H (2017). *TWOSEX-MSChart: A computer program for the age-stage, two-sex life table analysis* <http://140.120.197.173/Ecology>
- Chi H (1988). Life-table analysis incorporating both sexes and variable development rates among individuals. *Environ Entomol* 17:26–34
- hi H, H Liu (1985). Two new methods for the study of insect population ecology. *Bull Inst Zool Acad Sin* 24:225–240
- Cutler GC, CD Scott-Dupree, JH Tolman, CR Harris (2005). Acute and sublethal toxicity of novaluron, a novel chitin synthesis inhibitor, to *Leptinotarsa decemlineata* (Coleoptera: Chrysomelidae). *Pest Manage Sci* 61:1060–1068
- Desneux N, A Decourtye, JM Delpuech (2007). The sublethal effects of pesticides on beneficial arthropods. *Annu Rev Entomol* 52:81–106
- Goodman D (1982). Optimal life histories, optimal notation, and the value of reproductive value. *Amer Nat* 119:803–823
- Gulzar A, DJ Wright (2014). Fluctuating asymmetry, morphological changes and flight muscle ratio in a Vip3A resistant subpopulation of *Heliothis virescens* (WF06). *Intl J Agric Biol* 16:416–420
- Hafeez M, S Jan, M Nawaz, E Ali, B Ali, M Qasim, GM Fernandez-Grandon, M Shahid, M Wang (2019). Sub-lethal effects of lufenuron exposure on spotted bollworm *Earias vittella* (Fab): Key biological traits and detoxification enzymes activity. *Environ Sci Pollut Res* 26:14300–14312
- Han W, S Zhang, F Shen, M Liu, C Ren, X Gao (2012). Residual toxicity and sublethal effects of chlorantraniliprole on *Plutella xylostella* (Lepidoptera: Plutellidae). *Pest Manage Sci* 68:1184–1190
- Huang YB, H Chi (2013). Age-stage, two-sex life tables of *Bactrocera cucurbitae* (Coquillett) (Diptera: Tephritidae) with a discussion on the problem of applying female age-specific life tables to insect populations. *Ins Sci* 19:263–273
- Ishaaya I, A Barazani, S Kontsedalov, AR Horowitz (2007). Insecticides with novel modes of action: Mechanism, selectivity and cross-resistance. *Entomol Res* 37:148–152
- Ishaaya I, S Kontsedalov, AR Horowitz (2003). novaluron (Rimon), a novel IGR: Potency and cross-resistance. *Ins Biochem Physiol* 54:157–164
- Jan MT, N Abbas, SA Shad, MA Saleem (2015). Resistance to organophosphate, pyrethroid and biorational insecticides in populations of spotted bollworm, *Earias vittella* (Fabricius) (Lepidoptera: Noctuidae), in Pakistan. *Crop Prot* 78:247–252
- Lashkari MR, A Sahragard, M Ghadamyari, (2007). Sublethal effects of imidacloprid and pymetrozine on population growth parameters of cabbage aphid, *Brevicoryne brassicae* on rapeseed, *Brassica napus* L. *Ins Sci* 14:207–212
- Lohmeyer KH, JM Pound (2012). Laboratory evaluation of novaluron as a development site treatment for controlling larval horn flies, house flies, and stable flies (Diptera: Muscidae). *J Med Entomol* 49:647–651
- Meng QW, JJ Wang, JF Shi, WC Guo, GQ Li (2018). Effect of teflubenzuron ingestion on larval performance and chitin content in *Leptinotarsa decemlineata*. *Amer J Potato Res* 95:463–472
- Papachristos DP, PG Milonas (2008). Adverse effects of soil-applied insecticides on the predatory coccinellid *Hippodamia undecimnotata* (Coleoptera: Coccinellidae). *Biol Cont* 47:77–81
- Perveen F, T Miyata (2000). Effects of sublethal dose of chlorfluzuron on ovarian development and oogenesis in the common cutworm *Spodoptera litura* (Lepidoptera: Noctuidae). *Ann Entomol Soc Amer* 93:1131–1137
- Popp J, K Hantos (2011). The impact of crop protection on agricultural production. *Stud Agric Econ* 113:47–66
- Praveen K, AS Sajjan, R Patil, P Dharmatti, M Kurdikeri (2007). Influence of seed treatment and foliar spray with insecticides and neem products on growth and seed yield in okra (*Abelmoschus esculentus* [L] Moench). *Karnat J Agric Sci* 20:388–390
- Qu CW, F Zhang, G Li, CL Tetreau, R Wang (2017). Lethal and sublethal effects of dinotefuran on two invasive whiteflies, *Bemisia tabaci* (Hemiptera: Aleyrodidae). *J Asia Pac Entomol* 20:325–330
- R Development Core Team (2009). R: A language and environment for statistical computing. R Foundation for Statistical Computing, Vienna <http://www.R-project.org>
- Rahman MA, MM Uddin, MA Haque, MM Rahman (2016). Comparative efficacy of different tactics for the management of okra shoot and fruit borer, *Earias Vittella* (Fab.) under field condition. *Intl J Appl Sci Biotechnol* 4:74–78
- Rahmani S, AR Bandani (2013). Sublethal concentrations of thiamethoxam adversely affect life table parameters of the aphid predator, *Hippodamia variegata* (Goeze) (Coleoptera: Coccinellidae). *Crop Prot* 54:168–175
- Rasool B, J Arif, M Hamed, S Nadeem (2002). Field performance of *Trichogramma chilonis* against *Helicoverpa armigera* under varying sowing times and varieties of cotton. *Intl J Agric Biol* 4:113–114
- Rehan A, S Freed (2015). Fitness cost of methoxyfenozide and the effects of its sublethal doses on development, reproduction, and survival of *Spodoptera litura* (Fabricius) (Lepidoptera: Noctuidae). *Neotrop Entomol* 44:513–520
- Seth RK, JJ Kaur, DK Rao, SE Reynolds (2004). Effects of larval exposure to sublethal concentrations of the ecdysteroid agonists RH-5849 and tebufenozide (RH-5992) on male reproductive physiology in *Spodoptera litura*. *J Ins Physiol* 50:505–517
- Umrao RS, S Singh, J Kumar, D Singh, D Singh (2013). Efficacy of novel insecticides against shoot and fruit borer (*Earias vittella* Fabr.) in okra crop. *Hortic Flor Res Spectr* 2:251–254
- Yin XH, QJ Wu, XF Li, YJ Zhang, BY Xu (2008). Sublethal effects of spinosad on *Plutella xylostella* (Lepidoptera: Yponomeutidae). *Crop Prot* 27:1385–1391
- Zhang RM, EB Jang, SCJ He (2015). Lethal and sublethal effects of cyantraniliprole on *Bactrocera dorsalis* (Hendel) (Diptera: Tephritidae). *Pest Manage Sci* 71:250–256