# Full Length Article



# **Effects of Insoluble Fiber on Serum Biochemical Characteristics in Broiler**

M. SARIKHAN, H.A. SHAHRYAR<sup>1</sup>, K. NAZER-ADL, B. GHOLIZADEH AND B. BEHESHT Department of Animal Science, Islamic Azad University Shahbestar branch, Joy Park of Shabestar, Iran <sup>1</sup>Corresponding author's e-mail: h\_a\_shahriar@yahoo.com

## ABSTRACT

An experiment was conducted to determine the effects of insoluble raw fiber concentrate (IRFC) on blood biochemical values in broiler males. Four dietary treatments with three replicates containing 0, 0.25, 0.50 and 0.75% of IRFC were fed to 180 day old Lohmann chicks from 1 to 42 days. The chicks were randomly assigned to 12 cages (15 birds per cage). The experimental period was 42 days and serum biochemical values containing triglyceride, cholesterol, HDL, LDL, VLDL, total protein, albumin, amylase, lipase, calcium and phosphorous were measured at the end of starter (21 d) and grower (42 d) periods. Levels of total protein, albumin, amylase, lipase and phosphorous were not affected much by dietary treatments at day 42. Triglyceride, HDL and VLDL levels affected (p<0.01) by different levels of IRFC in diet and they were lower levels in triglyceride and VLDL and higher levels in HDL in 0.50 and 0.75 dietary groups than control and 0.25 groups, also cholesterol and LDL levels in 0.75 group were significantly lower (p<0.01) as compared to control group. Levels of total protein, albumin, amylase, lipase and phosphorous did not affected by dietary treatments but Ca in 0.50 group was higher than controls.

Key Words: Insoluble fiber; Serum biochemical; Diet; Broiler

## **INTRODUCTION**

Dietary fiber can be defined as component of plants, which is resistant to digestion by endogenous enzymes. The functional fiber refers to isolated, extracted, or synthetic fiber that has proven health benefits, with several physiological functions (Bersamin & Zidenberg-Cherr, 2004). Most insoluble fibers are moderately or slowly fermented. Those highly resistant to fermentation include isolated cellulose and lignin. Lignin is not fermented due to its composition as a phenyl propane polymer rather than carbohydrate (Whiteley *et al.*, 1996; Klurfeld, 1999). Carbohydrates common in poultry diets are starch, sugars, cellulose and other non-starch compounds. Cellulose and non-starch compounds are typically classified as crude fiber (Wilson & Beyer, 2000).

The major chemical component important to the structural integrity of grains is the insoluble fiber, which makes up the main part of the cell wall architecture. Indeed, insoluble fiber itself has shown beneficial effects on nutrient digestion (Svihus & Hetland, 2001; Hetland *et al.*, 2005). Dietary fiber (DF) content, have an important place in the well-balanced diets. Differentiation of water-soluble and insoluble fiber components has helped elucidate the physiological effects of fiber (Newman *et al.*, 1992).

Fiber is a nutritionally, chemically and physically heterogeneous material. This heterogeneous mix can be categorized into two major subclasses i.e., soluble, viscous and fermentable fiber (soluble) and insoluble, no viscous and no fermentable fiber (insoluble). The two subclasses have different roles in the digestive/absorptive processes within the gastrointestinal tract. The ratio of insoluble to soluble fiber (I: S) in a DF source can affect overall diet utilization and appears to be important in the formulation of diets to provide optimal efficacy (Burhalter *et al.*, 2001).

It is well established that the ingestion of some types of dietary influence lipid levels (Razdan & Pettersson, 1994; Durdi & Gharejeh, 2001). Structural differences between fibers have been reported (Woodward *et al.*, 1988; Jeraci & Lewis, 1999). These include differences in molecular weight (Cui, 2001) and solubility (Aman & Graham, 1987) but the cholesterol-lowering properties are approximately equivalent (Delaney *et al.*, 2003).

The non-digestible carbohydrates (dietary fiber) have been reported to improve the intestinal absorption of minerals, presumably because of their binding or sequestering action (Roberfroid *et al.*, 2002; Coudray *et al.*, 2003). Roberfroid (2000) indicated that a higher concentration of short-chain carboxylic acids resulted from the colonic fermentation of non-digestible carbohydrates, accelerating the colonic absorption of minerals, particularly calcium (Ca<sup>2+</sup>) and magnesium (Mg<sup>2+</sup>). The objective of this study was to determine the effect of different levels of IRFC on serum biochemical broiler.

# MATERIALS AND METHODS

Birds and diets. The experiment was carried out in a

To cite this paper: Sarikhan, M., H.A. Shahriyar, K. Nazer\_Adl, B. Gholizadeh and B. Beheshti, 2009. Effects of insoluble fiber on serum biochemical characteristics in broiler. *Int. J. Agric. Biol.*, 11: 73–76

completely randomized design using 180 days-old broiler chicks (Lohmann) were weighted and distributed randomly to 4 treatments with 3 replicates (15 chicks in each replicate per pen). Experimental diets, formulated according to NRC (1994), included following levels of Insoluble raw fiber concentrate (IRFC): a) control diet (no IRFC), b) 0.25%, c) 0.50% and d) 0.75% from Vitacel R200 (Table I). Birds were fed with experimental diet for starter (0-21 d) and grower (22-42 d) periods (Table I).

**Samples procedures.** At days 21 and 42, 2 chicks from each pen were selected and blood samples collected. The analysis of serum, triglyceride (TG), cholesterol (CHOL), high density lipoprotein (HDL), total protein (TP), albumin (ALB), amylase (AML), lipase (LIP),  $Ca^{2+}$  and Phosphorous (P), were measured on autoanalyzer (ALCYON 300-Abbott, USA) using commercially available kits. Low density lipoprotein cholesterol (LDL-C) and very low density lipoprotein cholesterol (VLDL-C) levels were estimated with Friedewald *et al.* (1972) equation.

**Statistical analysis.** Data were subjected to analysis of variance and significant differences observed in means subjected to Duncan's multiple range test. All data were analyzed for variance analysis using the general linear model (GLM) procedures of the SAS Institute (SAS Institute, 2003).

## **RESULTS AND DISCUSSION**

At day 21, serum TG, CHOL, HDL, LDL, VLDL levels were not influenced by different levels of IRFC in diet, but levels of all this parameters were significantly (p < p0.01) affected by levels of IRFC in dietary groups at day 42 (Table II). Serum TG levels were significantly lower in broilers fed diets containing 0.50% IRFC and 0.75% IRFC. Result showed that in 0.75% IRFC group a reduction in serum CHOL and LDL in compare with control group. Also higher concentrations of HDL and lower concentrations of VLDL observed in the serum of 0.50% IRFC and 0.75% IRFC dietary groups. Bile lipids compositions in mammals are mainly phospholipids and cholesterol but poultry contains cholesterol esters and triglycerides. According the Leeson et al. (1997) excretion of these lipids may be have a regulatory effects and this can describe the reduction of triglyceride levels with using IRFC due to ability of fibers to binding with lipid compositions.

A negative correlation exists between dietary fiber content and serum cholesterol level (Stasse Wolthuis *et al.*, 1980; Petterson & Aman, 1992; Sundberg *et al.*, 1995). Moundras *et al.* (1997) reported that the plasma cholesterol lowering effect of crude fiber may be due to its ability to enhance fecal excretion of cholesterol and bile acids. Daggy *et al.* (1997) reported that the fiber induces both enhanced

Ingredient and composition	Starter (0	-21 d)			Grower (2	Grower (22-42 d)			
	1	2	3	4	1	2	3	4	
Ground yellow corn	63.85	63.70	63.74	63.49	69.36	69.26	69.12	68.93	
Soybean meal (48% CP)	27.00	27.00	26.86	26.90	24.75	24.58	24.47	24.40	
Fish meal, menhaden (60% CP)	6.00	5.9	6.00	5.95	2.86	2.88	2.88	2.90	
DCP	1.00	1.00	0.82	0.84	0.80	0.80	0.80	0.80	
Oyster shell	1.20	1.20	1.13	1.12	1.32	1.32	1.32	1.32	
Sodium chloride	0.20	0.20	0.20	0.20	0.20	0.20	0.20	0.20	
Vitamin premix <sup>1</sup>	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	
Mineral premix <sup>2</sup>	0.25	0.25	0.25	0.25	0.25	0.25	0.25	0.25	
DL-methionine	0.20	0.20	0.20	0.20	0.10	0.10	0.10	0.10	
L-lysineHCL	0.00	0.00	0.00	0.00	0.06	0.06	0.06	0.05	
Coccidiostats	0.05	0.05	0.05	0.05	0.05	0.05	0.05	0.05	
IRFC <sup>3</sup>	0.00	0.25	0.50	0.75	0.00	0.25	0.50	0.75	
Calculated analysis									
ME, kcal/kg	2910	2900	2900	2895	2960	2950	2940	2935	
$CP(N \times 6.25)$	20.91	20.84	20.84	20.81	18.50	18.43	18.37	18.34	
ME/CP	139.1	139.1	139.1	139.1	160.0	160.0	160.0	160.0	
Crude fiber	3.34	3.51	3.69	3.84	3.28	3.45	3.62	3.79	
Ether extract	3.21	3.19	3.20	3.19	3.10	3.10	3.09	3.09	
Lysine	1.16	1.16	1.16	1.16	1.02	1.02	1.01	1.00	
TSAA	0.90	0.90	0.90	0.90	0.73	0.73	0.72	0.72	
Ca <sup>2+</sup>	1.07	1.06	1.00	1.00	0.9	0.9	0.9	0.9	
Available P	0.48	0.48	0.45	0.45	0.36	0.36	0.36	0.36	
Ca <sup>2+</sup> :P ratio	2.22	2.21	2.22	2.22	2.50	2.50	2.50	2.50	

 Table I. Composition of experimental diets (%)

<sup>1</sup> The vitamin premix supplied the following per kilogram of complete feed: vitamin A, 4,500 IU (retinyl acetate); cholecalciferol, 1,000 IU; vitamin E, 25 IU (dl-a-tocopheryl acetate); vitamin B12, 0.02 mg; menadione, 1.5 mg; riboflavin, 3 mg; thiamine, 1.5 mg; pantothenic acid, 5 mg; niacin, 20 mg; choline, 150 mg; folic acid, 0.5 mg; biotin, 0.5 mg; pyridoxine, 2.5 mg.

<sup>2</sup> The mineral premix supplied the following per kilogram of complete feed: manganese (MnSO4·H2O), 60 g; zinc (ZnO), 40 mg; iron (FeSO4·7H2O), 80 mg; copper (CuSO4·5H2O), 8 mg; selenium (Na2SeO3), 0.2 mg; iodine (Iodized NaCl), 0.8 mg; cobalt (CoCl2), 0.4 mg.

<sup>3</sup> IRFC: Insoluble raw fiber concentrate type Vitacel R200 is a commercial product of JRS Co. (Germany) with this composition, DM: 93.9%, crude fiber 72.5%, ADF: 86.5%, ADL: 0.6%, NDF: 90.5%, crude protein ( $n \times 6.25$ ): 1.1%, crude fat 0.2%, NFE: 19.8%, crude ash 0.3%, sugar: <0.1%, crude starch: <0.1%.

NS \*\*

Diet					Serun	n lipids <sup>1</sup>				
	T	riglyceride	С	holesterol		HDL		LDL		VLDL
	21 d	42 d	21 d	42 d	21 d	42 d	21 d	42 d	21 d	42 d
0.00% IRFC	90.8	106.6 <sup>a</sup>	144.6	143.5 <sup>a</sup>	32.0	33.8 <sup>b</sup>	94.5	88.5 <sup>a</sup>	18.1	21.3 <sup>a</sup>
0.25% IRFC	83.6	100.2 <sup>a</sup>	146.3	135.3 <sup>b</sup>	29.0	34.0 <sup>b</sup>	100.0	81.3 <sup>ab</sup>	16.7	20.1 <sup>a</sup>
0.50% IRFC	83.5	80.8 <sup>b</sup>	132.3	129.3 <sup>bc</sup>	32.0	38.4ª	83.6	74.8 <sup>bc</sup>	16.7	16.2 <sup>b</sup>
0.75% IRFC	83.3	75.8 <sup>b</sup>	134.6	123.3°	42.0	39.5ª	76.0	68.3°	16.6	15.2 <sup>b</sup>
SEM <sup>2</sup>	2.51	4.48	3.20	2.53	2.09	2.86	4.04	2.56	0.50	0.90
Source of Varian	ice				Probabili	ties				

Table II. Blood serum lipids of chickens fed different levels of IRFC (mg dL<sup>-1</sup>)

NS

NS  $^{1}$ n = 6 samples within each treatment group.

Diet

<sup>2</sup>SEM, based on pooled estimate of variance and n = 3.

<sup>a,b</sup>Means within columns with no common superscript differ significantly (P < 0.05) by Duncan's multiple range test.

#### Table III. Blood serum proteins of chickens fed different levels of IRFC

Table	V.	Blood	serum	minerals	of	chickens	fed
differe	nt le	vels of I	RFC				

\*\*

NS

Diet	Serum proteins <sup>1</sup>						
		proteir		Albumin			
	21 d	42 d		21 d	42 d		
			$(g dL^{-1})$				
0.00% IRFC	3.30	3.51		1.86	1.91		
0.25% IRFC	3.30	3.81		1.93	2.06		
0.50% IRFC	3.53	4.01		1.90	2.15		
0.75% IRFC	3.65	3.65		1.86	2.18		
SEM <sup>2</sup>	0.08	0.08		0.02	0.04		
Source of Variance			Probabilities				
Diet	NS	NS		NS	NS		

 $^{1}$ n = 6 samples within each treatment group.

<sup>2</sup>SEM, based on pooled estimate of variance and n = 3.

<sup>a,b</sup>Means within columns with no common superscript differ significantly (P < 0.05)

Table IV. Blood serum enzymes of chickens fed different levels of IRFC

Diet	Serum enzymes <sup>1</sup>							
	Ar	nylase		Lipase				
	21 d	42 d		21 d	42 d			
			(µg L <sup>-1</sup> )					
0.00% IRFC	754.7	1270.0		129.3	181.1			
0.25% IRFC	803.7	1221.5		149.7	192.0			
0.50% IRFC	904.0	1346.3		153.3	188.2			
0.75% IRFC	759.0	938.5		135.3	182.5			
SEM <sup>2</sup>	36.59	79.23		4.39	7.31			
Source of Variance			Probabilities					
Diet	NS	NS		NS	NS			

 $^{1}$ n = 6 samples within each treatment group.

<sup>2</sup>SEM, based on pooled estimate of variance and n = 3.

<sup>a,b</sup>Means within columns with no common superscript differ significantly (P < 0.05)

liver excretion and diversion of intestinal steroids to the feces. Durdi and Gharejeh (2001) reported that reduction in total cholesterol concentration and increased HDL to total cholesterol ratio is probably cause by enhanced reverse cholesterol transport in response to intestinal loss of dietary fat. Mathlouthi et al. (2002) reported that indigestible polysaccharides can act directly by increasing bile acid excretion. Garcia-Diez (1996) and Adrizal and Ohtani (2002) confirmed that non-starch polysaccharides have

Diet	Serum	ı mineral				
	Ca	lcium		Pho	Phosphorous	
	21 d	42 d		21 d	42 d	
			$(mg dL^{-1})$			
0.00% IRFC	9.40	8.63 °		5.63	5.15	
0.25% IRFC	9.60	8.68 bc		5.16	5.18	
0.50% IRFC	9.60	9.23 <sup>a</sup>		5.53	5.91	
0.75% IRFC	10.13	9.13 <sup>ab</sup>		5.16	5.66	
SEM <sup>2</sup>	0.14	0.10		0.18	0.13	
Source of Variance			Probabilities			
Diet	NS	*		NS	NS	

 $^{1}$ n = 6 samples within each treatment group.

NS

<sup>2</sup>SEM, based on pooled estimate of variance and n = 3.

<sup>a,b</sup>Means within columns with no common superscript differ significantly (P < 0.05)

binding property with bile acids. This results in increased fecal and reduced serum cholesterol. Reduction in cholesterol parameter cause increase in HDL and reduce LDL and VLDL in serum. Serum concentrations of total protein (TP), albumin (AL) (Table III) and levels of amylase (AML) and Lipase (LIP) in serum (Table IV) had no significant differences (p > 0.05) at days 21 and 42. Levels of phosphorous in serum were not affected (P > 0.05) by dietary treatments at days 21 and 42. Besides this, serum Ca<sup>2+</sup> concentration did not show any differences at day 21 but it was significantly (P< 0.05) affected at day 42 (Table V). At day 42 birds fed 0.50% IRFC in diet had higher levels of  $Ca^{2+}$  in serum compared with controls. In concurrence with these findings, Roberfroid (2000), Coudray et al. (2003) and Chen and Chen (2004) reported that a higher concentration of short-chain carboxylic acids resulting from the colonic fermentation of non-digestible carbohydrates accelerates the colonic absorption of minerals, particularly Ca2+ and Mg2+. It seems that this procedure help increasing Ca<sup>2+</sup> levels in serum.

#### **CONCLUSION**

Increase of dietary fiber level resulted in improves the performance and reduces TG, CHOL, LDL, VLDL concentrations in serum of broiler.

Acknowledgment. Financial supports for this study was provided by Islamic Azad University, Shabestar Branch and JRS Company from Germany. The authors thank A.M. Vatankhah for his skilled technical assistance throughout the experimental analyses.

## REFERENCES

- Adrizal, O. and S. Ohtani, 2002. Defatted rice bran non starch polysaccharides in broiler diets: Effects of supplements on nutrient digestibilities. J. Poult. Sci., 39: 67–76
- Aman, P. and H. Graham, 1987. Analysis of total and insoluble mixedlinked (1→3) (1→4)-β-D-glucanas in barley and oats. J. Agric. Food Chem., 35: 704–9
- Burhalter, T.M., N.R. Merchen, L.L. Bauer, S.M. Murray, A.R. Patil, J.L. Brent and G.C. Fahey, 2001. The ratio of insoluble to soluble fiber components in soybean hulls affect ileal and total-tract nutrient digestibilities and fecal characteristics of dogs. J. Nutr., 131: 1978– 1985
- Bersamin, A. and S. Zidenberg-Cherr, 2004. *Nutrition and Health, Some Facts about Fiber*. Department of nutrition university of California Davis. Division of Agriculture and Natural Resources. An electronic version of this publication is available on the ANR Communication Services Web site at: http://anrcatalog.ucdavis.edu
- Chen, Y.C. and T.C. Chen, 2004. Mineral utilization in layers as influenced by dietary oligofructose and inulin. *Int. J. Poult. Sci.*, 3: 442–445
- Coudray, C., J.C. Tressol, E. Gueux and Y. Rayssiguier, 2003. Effects of inulin-type fructans of different chain length and type of branching on intestinal absorption and balance of calcium and magnesium in rats. *European J. Nutr.*, 42: 91–98
- Cui, S.W., 2001. Cereal non-starch polysaccharides: (1→3) (1→4)-β-Dglucanas. In: Polysaccharide from Agricultural Products Processing, Structures and Functionality, pp: 103–166. Technomic Publishing Lancater, PA
- Daggy, B.P., N.C. O'Connell, G.R. Jerdak, B.A. Stinson and K.D.R. Setchell, 1997. Additive hypocholesterolemic effect of psyllium and cholestyramine in the hamster: influence on fecal sterol and bile acid profiles. J. Lipid. Res., 38: 491–502
- Delaney, B., R.J. Nicolosi, T.A. Wilson, T. Carison, F. Frazer, G.H. Zheng, R. Hess, K. Ostergren. J. Haworth and N. Knutson, 2003. β-glucan fractions from barley and oats are similarly antiatherogenic in hypercholesterolemia Syrian golden hamsters. J. Nutr., 133: 468–495
- Durdi, Q. and A.M. Gharejeh, 2001. Effects of dietary chitosan on nitrogen metabolite levels in mice. Arch. Irn. Med., 4: 96–98
- Friedewald, W.T., R.I. Levy and D.S. Fredrickson, 1972. Estimation of concentration of low-density lipoprotein cholesterol in plasma without use of the ultra-centrifuge. *Clin. Chem.*, 18: 449–502
- Garcia-Diez, F., V. Garcia-Mediavilla, J.E. Bayon and J. Gonzalez-Gallego, 1996. Pectin feeding influences fecal bile acid excretion, hepatic bile acid and cholesterol synthesis and serum cholesterol in rats. J. Nutr., 126: 1766–1771
- Hetland, H., B. Svihus and M. Choct, 2005. Role of insoluble fiber on gizzard activity in layers. J. Appl. Poult. Res., 14: 38–46
- Jeraci, J.L. and B.A. Lewis, 1999. Detrmination of soluble fiber components: (1→3: 1→4)-β-D-glucanas and pectins. *Anim. Feed Sci. Technol.*, 23: 15–25

- Klurfeld, D.M., 1999. Nutritional regulation of gastrointestinal growth. *Frontiers Biosci.*, 4: 299–302
- Leeson, S., A.K. Zubair, E.J. Squires and C. Forsberg, 1997. Influence of dietary levels of fat, fiber and copper sulfate and fat rancidity on cecal activity in the growing turkey. *Poult. Sci.*, 76: 59–66
- Mathlouthi, N., J.P. Lalles, P. Lepersq, C. Juste and M. Larbier, 2002. Xylanase and  $\beta$ -glucanase supplementation improve conjugated bile acid fraction in intestinal contents and increase villus size of small intestine wall in broiler chickens fed ray-based diet. *J. Anim. Sci.*, 80: 2773–2779
- Moundras, C., S.R. Behr, C. Remesy and C. Demigne, 1997. Fecal losses of sterols and bile acids induced by feeding rat's guar gum are due to greater pool size and liver bile acid secretion. J. Nutr., 127: 1068–76
- Newman, R.K., C.F. Klopfensten, C.W. Newman, N. Guritno and P.J. Hofer, 1992. Comparison of the cholesterol-lowering properties of whole barley, oat bran and red dog in chicks and rats. *Cereal Chem.*, 69: 240–244
- NRC, 1994. Nutrient Requirements of Poultry: Nutritional Requirements of Poultry, 9<sup>th</sup> edition. National Academy of Science, Washington, DC
- Petterson, D. and P. Aman, 1992. Production responses and serum lipid concentration of broiler chickens fed diets based on oat bran and extracted oat bran with and without enzyme supplementation. J. Sci. Food Agric., 58: 569–576
- Razdan, A. and D. Pettersson, 1994. Effect of chitin and chitosan on nutrient digestibility and plasma lipid concentration in broiler chickens. *British J. Nutr.*, 72: 277–288
- Roberfroid, M.B., 2000. Prebiotics and probiotics: are they functional foods? *American J. Clin. Nutr.*, 71 (Suppl): 1682S–7S
- Roberfroid, M.B., J. Cumps and J.P. Devogelaer, 2002. Dietary chicory inulin increases whole-body bone mineral density in growing male rats. J. Nutr., 132: 3599–3602
- SAS Institute, 2002-2003. SAS User's Guide: Statistics, Version 9.1, edition. SAS Institute Inc. Cary, NC, 27513, USA
- Stasse Wolthuis, M., H.F.F. Albers., J.G.C. Vanjaveren, J. Wildejong, J.G.A.J. Hautvest, R.J.J. Hermus, M.B. Katan, W. Gordon Brydon and M.A. Eastwood, 1980. Influence of dietary fiber from vegetables and fruits, bran or citrus pectin on serum lipids, fecal lipids and colonic function. *American J. Clinical Nutr.*, 33: 1745–1756
- Sundberg, B., D. Petterson and P. Aman, 1995. Nutritional properties of fiber rich barley products fed to broiler chickens. J. Sci. Food Agric., 67: 469–476
- Svihus, B. and H. Hetland, 2001. Ileal starch digestibility in growing broiler chickens fed a wheat-based diet is improved by mash feeding, dilution with cellulose or whole wheat inclusion. *British Poult. Sci.*, 42: 633–637
- Whiteley, L.O., M.P. Purdon, G.M. Ridder and T.A. Bertram, 1996. The interactions of diet and colonic microflora in regulating colonic mucosal growth. *Toxicol Pathol.*, 24: 305–314
- Wilson, K.J. and R.S. Beyer, 2000. *Poultry Nutrition Information for Small Flock*. Publication from Kansas State University, available on the World Wide Web at: http://www.oznet.ksu.edu
- Woodward, J.R., D.R. Philips and G.B. Fincher, 1988. Water-soluble (1→3, 1→4)-β-D-glucanas from barley (*Hordeum vulgare*) endosperm. I. Physicochemical properties. *Carbohydrate Polym.*, 3: 143

#### (Received 29 April 2008; Accepted 29 August 2008)