Full Length Article



Determination of Different Geographical Populations of *Rhynchophorus ferrugineus* (Olivier) (Coleoptera: Curculionidae) using RAPD-PCR

RABAB A.A.M. EL-MERGAWY¹[†], ABDULAZIZ M. AL AJLAN[‡], NAGLAA A. ABDALLAH[¶], MAHMOUD I. NASR[†] AND JEAN-FRANÇOIS SILVAIN

IRD, UR 072, Laboratoire Evolution, Génomes et Spéciation, UPR 9034, CNRS, 91198 Gif-sur-Yvette, France

[†]Department of Molecular Biology, Genetic Engineering and Biotechnology Research Institute (GEBRI), Minoufyia University, El-Sadat City, Minoufyia, Egypt

[‡]Department of Arid Land Agriculture, College of Agriculture and Food Sciences, King Faisal University, PO Box 55009, Hofuf 31982, Kingdom of Saudi Arabia

¶Department of Genetics, Faculty of Agriculture, Cairo University, Egypt

¹Corresponding author's e-mail: rababml@yahoo.fr

ABSTRACT

This study investigated the genetic variation among thirteen geographic populations of the red palm weevil (RPW), *Rhynchophorus ferrugineus* (Olivier), using the Random Amplified Polymorphic DNA-Polymerase Chain Reaction (RAPD-PCR) technique. RAPD profiles exhibited bands between 200 to 1400 bp in length. Average genetic distances among populations ranged from 0.3 to 0.8. Genetic distances among the analyzed individuals ranged from 0.241 (between individuals from populations of Egypt & Turkey) to 1 (between individuals from populations of Japan & both Egypt, Spain & France). The Unweigthed Pair Group Method with Arithmetic (UPGMA) mean categorized the tested populations into twelve clusters. Based on results, it is suggested that there are two major routes of RPW invasion, the first is at the origin of all the populations found in Egypt and Mediterranean countries and second is at the origin of the populations found in the Arabic Peninsula, Iran and Pakistan. © 2011 Friends Science Publishers

Key Words: RAPD-PCR; Rhynchophorus ferrugineus; Invasive species; DNA fingerprinting

INTRODUCTION

The Red Palm Weevil (RPW), Rhynchophorus ferrugineus (Olivier) (Coleoptera: Curculionidae), is an invasive species that is claimed to be originated from South and Southeast of Asia, where it is considered as one of the most destructive pests of coconut, Cocos nucifera L. (Abraham et al., 1975; Sivapragasam et al., 1990; Sadakathulla, 1991; Murphy & Briscoe, 1999; Ferry & Gomez, 2002). The invasion history of RPW to the Middle East and the Mediterranean area date back to the mid 80's where it was detected for the first time in the United Arab Emirates (UAE), since then the pest is continuing its invasion to the other countries of Middle-East, including Egypt, and to most of the countries of the Northern shore of the Mediterranean sea up to Portugal (Cox, 1993; Barranco & Cabellot, 1996; Esteban-Durán et al., 1998; Kehat, 1999; Karut & Kazak, 2005; Kontodimas et al., 2007; EPPO, 2008; Ministry for Resources & Rural Affairs of Malta 2008; EPPO Reporting Service 2008, 2009). In those invaded countries RPW is threatening a broad host range of palms

and is considered as the most destructive pest of palms, where it spends its life cycle inside the trunk without visible sign (Nirula, 1956; Wattanapongsiri, 1966; Abraham *et al.*, 1998; Kaakeh, 2005; Faleiro, 2006a, b). The infestation of this insect is detected when trees are extensively damaged and it is not possible to recover (Gadelhak & Enan, 2005).

In a given invaded country, we can expect that the insect genetic diversity will be similar in case of a single introduction of insects as well as multiple successive introductions from the same source area. On the contrary multiple introductions from different source areas will likely increase the genetic diversity in the populations as they introduce new alleles (Grapputo *et al.*, 2005).

Considering the economic importance of RPW, it is necessary to know the history of its invasion and to understand the relationships between the different geographic populations. Studying the genetic diversity of RPW will help answering several questions such as: the number of invasion routes from Asia to the West and the number of introductions that have occurred in each of the invaded countries. In addition, this study of genetic variation

To cite this paper: El-Mergawy, R.A.A.M., A.M. Al Ajlan, N.A. Abdallah, M.I. Nasr and J.F. Silvain, 2011. Determination of different geographical populations of *Rhynchophorus ferrugineus* (olivier) (coleoptera: curculionidae) using RAPD-PCR. *Int. J. Agric. Biol.*, 13: 227–232

among the geographic populations of invasive species is essential for designing their management strategy including biosecurity as it gives rapid and accurate identification of alien species and an accurate characterization of their populations (Armstrong & Ball, 2005; Grapputo *et al.*, 2005; Marimuthu *et al.*, 2009; Sharma *et al.*, 2009).

RAPD technique is considered as an useful tool as it need tiny amounts of DNA to give rapid and accurate identification of alien species especially in the developing countries, where DNA-based methods are unavailable due to their high cost, the requirement for complex equipments and the requirement for expertise (Williams *et al.*, 1990; Hadrys *et al.*, 1992; Bardakci, 2000).

The genetic variation of RPW was previously detected using RAPD markers. Those studies were restricted to comparisons among seven individuals of RPW from UAE, individuals of RPW from Egypt, KSA and Indonesia and different morphological forms of RPW individuals from Egypt and KSA (Abulyazid *et al.*, 2002; Salama & Saker, 2002; Gadelhak & Enan, 2005; Al-Ayied *et al.*, 2006).

In the present study RAPD was used to investigate the genetic variation and relationships among and within RPW from thirteen different countries.

MATERIALS AND METHODS

RPW samples: Samples of RPW were collected from 27 localities belonging to thirteen infested countries (Table I). In the present study, the comparison was done at both the population and individual levels. Population signified one or more individuals from the same area, while individual referred to a single individual.

DNA extraction: Total genomic DNA was extracted from RPW samples using DNeasy Tissue Kit (Qiagen GmbH) according to the manufacturer's protocol.

PCR amplification and electrophoresis of PCR products: Six different 10 mers random primers were used for the amplification of PCR products. Sequences of these primers and protocols used for DNA amplification and electrophoresis are the same as Gadelhak and Enan (2005).

Scoring bands and the binary matrix: Amplified products which were bright, reproducible and consistent in performance were chosen for scoring of shared and unshared bands. The bands were scored as present and absent where the presence of a particular band was denoted as 1 and absence as 0.

Similarity matrix: The binary matrix was subjected to the MSVP software ver. 3.12 d (Kovach Computing Services, Wales, UK) in order to detect the genetic similarity coefficient using the Nei and Li coefficient (Nei & Li, 1979) method.

Genetic distances among RPW individuals: The genetic distances were estimated to investigate the level of DNA variation among all the tested individuals. Each individual was treated as an Operation Taxonomic Unit (OTU). A matrix of genetic distances was calculated from the

similarity matrix according to Abulyazid *et al.* (2002) as follows: 1-similarity index.

Average genetic distances among the geographic populations: Average genetic distances were calculated with MEGA 4 (Tamura *et al.*, 2007) in order to compare among the thirteen geographic populations of RPW (Table I). The level of DNA variation among the Egyptian local populations was also calculated. The Egyptian individuals were divided into fifteen different populations according to their locality of origin (Table I).

Cluster analyses: Cluster analysis was performed based on the genetic distance matrix by means of the Unweighted Pair Group Method with Arithmetic mean (UPGMA) (Sneath & Sokal, 1973) with MEGA 4 (Tamura *et al.*, 2007).

RESULTS

Scored and polymorphic bands: The genetic variation among and within 13 geographic populations of RPW (Table I) was investigated using RAPD technique. The obtained RAPD profiles exhibited bands ranged from 200 to 1400 bp in length (Table II). All of the detected bands were polymorphic. The comparisons among RAPD profiles revealed that 17 RAPD markers were unique for the Egyptian local populations.

Comparison among populations: The average genetic distances were estimated to investigate the level of DNA variation among the 13 geographic populations of RPW (Table III). The lowest average genetic distance (0.3) was found between populations from KSA and UAE (0.268). The highest average genetic distance (approximately 0.8) was found between populations from KSA and Japan (0.759).

Average genetic distances among the fifteen local populations from Egypt: The average genetic distance among the 15 local populations from Egypt (Table I) ranged from 0.2 to 0.7 (Table IV).

Comparison among individuals: the genetic distances among the tested individuals were estimated. The lowest genetic distance 0.241 was found between the individuals Bur Said1 and Turkey2, while the highest genetic distance 1 was observed between Al Minufiyah2 and Japan1. The genetic distances between individuals from the same country ranged from 0.154 between the two individuals of Cyprus to 1 between the two individuals of Japan. Regarding the Egyptian individuals, the lowest genetic distance 0.158 was found between AlJizah2 and AlIsmailiyah2. The highest genetic distance 0.786 was found between AlFayyum2 and Dumyat1.

Cluster analyses: The UPGMA dendrogram showed 12 clusters, out of which six clusters included the tested individuals from Egypt (clusters 1 to 5 & 9), one of these six clusters (cluster 9) contained in addition to Egyptian individuals (Al Bahr al Ahmar & Al Buhayrah) the two tested ones from France (Fig. 1).

Geog	graphic	I	ndividual codes
popu	lations		
1	KSA		KSA 1&2
2	UAE		UAE1&2
3	Oman		Oman1&2
4	Turkey		Turkey1&2
5	Pakistan		Pakistan1&2
6	Iran		Iran1&2
7	Japan		Japan1&2
8	Spain		Spain1&2
9	Italy		Italy1&2
10	Greece		Greece1&2
11	Cyprus		Cyprus1&2
12	Franace		France1&2
13	Egypt	Egyptian local populations	
		Al Bahr alAhmar	AlBahralAhmar
			1&2
		Al Buhayrah	AlBuhayrah1&2
		Al Fayyum	AlFayyum1&2
		Iskandariyah	Iskandariyah1&
			2
		Al Ismailiyah	AlIsmailiyah1&
			2
		Al Jizah	AlJizah1&2
		Al Minufiyah	AlMinufiyah1&
			2
		Al Qalyubiyahg	AlQalyubiyahg
		Al WadialJadid	AlWadialJadid1
			&2
		Ash Sharqiyah	AshSharqiyah1
			&2
		Aswan	Aswan
		Beni Suwayf	BeniSuwayf1&2
		Bur Said	BurSaid1&2
		Dumyat	Dumyat1&2
		Kafr ashShykh	KafrashShykh1
		-	&2

Table I: Geographic populations and individual codes of RPW samples

Fig. 1: Dendrogram of the thirteen geographic populations of RPW derived from the genetic distances



DISCUSSION

Genetic variation among 13 geographic populations of RPW was detected using RAPD. Previous studies have demonstrated genetic variation of RPW from different geographic areas using RAPD (Abulyazid et al., 2002; Gadelhak & Enan, 2005). As well high genetic variation was detected among different geographical populations of other insect species using the same technique such as: Bombyx mori (Srivastava et al., 2005); Nebria gregaria (Clarke et al., 2001); Culex quinquefasciatus (Diptera: Culicidae) (Sharma et al., 2009); Aedes aegypti (Hiragi et al., 2009); Trypanosoma cruzi (Dib et al., 2009; Lala et al., 2009). In contrast, Marimuthu et al. (2009) found no genetic variation among different populations of shoot and fruit borer, Leucinodes orbonalis (Pyraustidae: Lepidoptera). This lack of genetic variation was explained by continuous gene flow between the different geographic populations.

An observed 100% polymorphism in RAPD markers has also been detected previously in the study of *Culex quinquefasciatus* (Diptera: Culicidae). This percentage reflects the absence of genetic homogeneity among the examined populations (Sharma *et al.*, 2009). In contrast Gadelhak and Enan (2005) detected 51.4% polymorphism in RAPD markers for comparison among seven RPW individuals from UAE. The unique RAPD markers observed in the present study may be used to produce genetic markers that can distinguish the geographic populations of RPW from each other (Haymer & McInnis, 1994; Bardakci, 2000).

The average genetic distance among the thirteen geographic populations of RPW ranged from 0.3 to 0.8. Similarly, a high average genetic distance among different geographic populations was detected in *Bombyx mori* (0.140-0.513) (Srivastava *et al.*, 2005). The highest and lowest average genetic distances revealed that similarity among the analyzed populations ranged from 20% to 70%. Similarity ranges from 80 to 100% and 38 to 94% were observed previously among whitefly species and RPW populations from UAE respectively (Perring *et al.*, 1993; Gadelhak & Enan, 2005).

High values of genetic distances among the 61 individuals of RPW were observed as the genetic distance between the analyzed individuals ranged from 0.241 to 1. Similarly, high genetic distances were observed previously among individuals of RPW from UAE (Gadelhak & Enan, 2005). Also, high genetic distances (0.058-0.513) were detected in *Bombyx mori* (Srivastava *et al.*, 2005) and *Chironomus plumosus* (Diptera: Chironomidae) (Gunderina *et al.*, 2009). According to the observed genetic distances among the tested individuals there is a positive correlation between the genetic distances and the geographical distances.

RAPD partitioned RPW populations from different geographic origins as the tested populations were grouped in

Primers	Number of bands													
	Egypt	KSA	UAE	Oman	Turkey	Iran	Pakistan	Japan	Spain	Italy	Greece	Cyprus	France	Total
Primer 1	100	5	6	6	4	5	6	4	2	6	5	5	5	159
Primer 2	69	2	7	2	3	8	7	2	2	2	7	2	3	116
Primer 3	74	8	5	5	3	7	4	3	5	5	3	4	4	130
Primer 4	59	2	3	3	4	6	4	4	3	3	3	3	4	101
Primer 5	68	4	4	4	6	6	7	4	5	5	5	6	4	128
Primer 6	87	4	4	3	5	4	4	4	2	8	7	6	4	142
Total	457	25	29	23	25	36	32	21	19	29	30	26	24	776

Table II: Number of bands calculated from 13 geographic populations (61 individuals) of RPW

Table III: Average genetic distances among the 13 RPW geographic populations

Populations	KSA	UAE	Oman	Turkey	Pakistan	Iran	Japan	Spain	Italy	Greece	Cyprus	France
KSA												
UAE	0.268											
Oman	0.433	0.273										
Turkey	0.650	0.673	0.542									
Pakistan	0.607	0.607	0.590	0.468								
Iran	0.475	0.418	0.513	0.708	0.311							
Japan	0.759	0.731	0.685	0.649	0.704	0.576						
Spain	0.678	0.732	0.716	0.759	0.641	0.677	0.708					
Italy	0.643	0.689	0.659	0.649	0.662	0.665	0.810	0.479				
Greece	0.593	0.671	0.612	0.540	0.588	0.629	0.787	0.511	0.377			
Cyprus	0.707	0.666	0.647	0.627	0.700	0.659	0.815	0.653	0.691	0.345		
France	0.662	0.667	0.600	0.579	0.667	0.594	0.833	0.540	0.538	0.577	0.348	
Egypt	0.561	0.603	0.626	0.600	0.612	0.624	0.758	0.639	0.552	0.602	0.555	0.632

Table IV: Average genetic distances among the 15 Egyptian local populations of RPW

Egyptian local populations				-	-		-							
	ь <u>н</u>	rah	E	yah	yah	ч	yah	ahg	did	ah	_	ayf	q	÷
	3ah ma	lay	лл.	lari	aili	iza	ilin	biy	vl lJa	sh qiy	wan	MI	Sai	nya
	AN I	Bul	Fa	and	Sm	L IV	Mir	lyu.≜	A	A	Asr	ni S	II.	III (
	B	II	I	Iska	IIV	~	N	Qal	Wa	\mathbf{S}		Bei	m	Π
Al Buhayrah	0.349													
Al Fayyum	0.575	0.367												
Iskandariyah	0.516	0.502	0.343											
Al Ismailiyah	0.502	0.476	0.413	0.340										
Al Jizah	0.473	0.470	0.440	0.386	0.246									
Al Minufiyah	0.519	0.535	0.524	0.459	0.369	0.368								
Al Qalyubiyahg	0.628	0.457	0.522	0.436	0.535	0.485	0.300							
Al WadialJadid	0.585	0.624	0.581	0.434	0.461	0.508	0.446	0.195						
Ash Sharqiyah	0.570	0.661	0.629	0.474	0.546	0.529	0.535	0.480	0.372					
Aswan	0.640	0.738	0.651	0.527	0.500	0.581	0.540	0.394	0.449	0.209				
Beni Suwayf	0.540	0.724	0.583	0.435	0.542	0.580	0.591	0.371	0.497	0.288	0.156			
Bur Said	0.591	0.674	0.592	0.496	0.513	0.579	0.576	0.404	0.519	0.352	0.321	0.302		
Dumyat	0.540	0.659	0.616	0.476	0.441	0.462	0.545	0.455	0.562	0.426	0.500	0.450	0.284	
Kafr ashShykh	0.511	0.612	0.522	0.372	0.447	0.470	0.477	0.277	0.420	0.358	0.333	0.377	0.325	0.221

12 clusters (Fig. 1). The UPGMA dendrogram clearly showed three groupings: (1) the Japanese population, (2) the Egyptian and Mediterranean populations and (3) Arabic Peninsula, Iran and Pakistan populations. This result was fairly important, because it is suggests that Egyptian and related Mediterranean populations have a different origin than Arabic Peninsula and Asian populations.

According to the dendrogram, not all the Egyptian individuals have direct relationships with local geographic pattern as some individuals from distant localities were clustered together: Cluster (2) Ash Sharqiyah, Aswan and Bani Suwayf; Cluster (3) Al Qalyubiyah, Al Wadi al Jadid and Ash Sharqiyah; Cluster (4) Al Fayyum and Iskandariyah (Fig. 1). A significant positive correlation between genetic variation and geographical distance was found for boll weevil *Anthonomus grandis* (Coleoptera: Curculionidae) populations in the United States (Kim & Sappington, 2004). In contrast, Gadelhak and Enan (2005) observed that the genetic and geographical distances among RPW populations from UAE were not correlated.

Invasive populations derived from multiple introductions from different origins are expected to be genetically more diverse (Vieira *et al.*, 2007). The UPGMA analysis suggested that multiple introductions from different countries or a single introduction composed of genetically diverse individuals may have occurred in Egypt as the Egyptian RPW individuals were clustered in six separate clusters (1, 2, 3, 4, 5, & 9) (Fig. 1), as well, the high genetic variability found within the Egyptian population (0.458) is supporting that finding.

In conclusion, our findings confirm that RAPD has the ability to distinguish among populations of RPW. The dendrogram and average genetic distances suggest that the RPW populations that invaded the Middle East and the Mediterranean area likely came from two different geographical origins. One is the source of the Egyptian and related Mediterranean populations and the other is the source of the Arabic Peninsula and Asian populations.

Acknowledgement: Thanks to all the colleagues who provided RPW samples: Santi Longo, Raffaele Griffo, Tomas Cabello Garcia, Eric Chapin, Laurence Olivier, J.P. Morin, Cengiz Kazak, Dimitris Kontodimas, Vassilis Vassiliou, Didier Rochat and Armand Faghih.

REFERENCES

- Abraham, V.A., K.M. Abdulla and C.H. Kurian, 1975. Evaluation of seven insecticides for control of red palm weevil *Rhynchophorus ferrugineus* Fabr. J. Plants Crops, 3: 71–72
- Abraham, V.A., M.A. Al-Shuaibi, J.R. Faleiro, R.A. Abozuhairah and P.S.P.V. Vidyasagar, 1998. An integrated management approach for red palm weevil *Rhynchophorus ferrugineus* Oliv. a key pest of date palm in the Middle East. *Agric. Sci.*, 3: 77–83
- Abulyazid, I., I.K.E. Kamel, F.A. Sharawi and S. El-Bermawi, 2002. Comparison between different populations of Red Palm Weevils *Rhynchophorus* species using RAPD-PCR. J. Egypt Ger. Soc. Zool., 38: 1–15
- Al-Ayied, H.Y., A.M. Alswailem, O. Shair and A.M. Al Jabr, 2006. Evaluation of phylogenetic between three phenotypically different forms of red date palm weevil *Rhynchophorus ferrugineus* Oliv. using PCR-based RAPD technique. *Arch. Phytopathol. Plant Prot.*, 39: 303–309
- Armstrong, K.F. and S.L. Ball, 2005. DNA barcodes for biosecurity: invasive species identification. *Philos. T. Roy. Soc. B.*, 360: 1813– 1823
- Bardakci, F., 2000. Random amplified polymorphic DNA (RAPD) markers. Turkish J. Biol., 25: 185–196
- Barranco, P.P.J. and T. Cabellot, 1996. El picudo rojo de las palmeras, *Rhynchophorus ferrugineus* (Olivier), nueva plaga en Europa. (Coleoptera, curculionidae). *Phytoma-España*, 76: 36–40
- Clarke, T.E., D.B. Levin, D.H. Kavanaugh and T.E. Reimchen, 2001. Rapid evolution in *Nebria gregaria* group (Coleoptera: Carabidae) and the paleogeography of the Queen Charlotte Islands. *Evolution*, 55: 1408– 1418
- Cox, M.L., 1993. Red palm weevil, Rhynchophorus ferrugineus in Egypt. FAO Plant Protect. B., 41: 30–31
- Dib J., C. Barnabé, M. Tibayrenc and O. Triana, 2009. Incrimination of *Eratyrus cuspidatus* (Stal) in the transmission of Chagas' disease by molecular epidemiology analysis of *Trypanosoma cruzi* isolates from a geographically restricted area in the north of Colombia. *Acta Trop.*, 111: 237–242
- EPPO (European and Mediterranean Plant Protection Organization), 2008. Datasheets on quarantine pests, *Rhynchophorus ferrugineus*. EPPO Bull., 38: 55–59
- EPPO (European and Mediterranean Plant Protection Organization) Reporting Service, 2008. First Report of Rhynchophorus ferrugineus in Portugal. 2008/022, No 2
- EPPO (European and Mediterranean Plant Protection Organization) Reporting Service, 2009. First Report of Rhynchophorus ferrugineus in Albania. 2009/207, No 11

- Esteban-Durán, J., J.L. Yela, F. Beitia-Crespo and A. Jiménez-Alvarez, 1998. Curculiónidos exóticos susceptibles de ser introducidos en España y otros países de la Unión Europa a través de vegetales importados (Coleoptera: Curculionidae: Rhynchophorinae). *Bol. San. Veg. Plagas*, 24: 23–40
- Faleiro, J.R., 2006a. A review of the Issues and management of the red palm weevil *Rhynchophorus ferrugineus* (Coleoptera: Rhynchophoridae) in coconut and date palm during the last one hundred years. *Int. J. Trop. Insect Sci.*, 26: 135–154
- Faleiro, J.R., 2006b. Insight into the Management of Red Palm Weevil: Based on Experiences on Coconut in India and Date Palm in Saudi Arabia. In: Agroalimed, F. (ed.), I Jornada Internacional Sobre el Picudo Rojo de las Palmeras (November, 2005), pp: 35–57. Fundacion Agroalimed. Valencia, Spain
- Ferry, M. and S. Gomez, 2002. The Red Palm Weevil in the Mediterranean Area. Palms, 46: 172–178
- Gadelhak, G.G. and M.R. Enan, 2005. Genetic diversity among populations of red palm weevil, *Rhynchophorus ferrugineus* Olivier (Coleoptera: Curculionidae), determined by random amplified polymorphic DNApolymerase chain reaction (RAPD-PCR). *Int. J. Agric. Biol.*, 7: 395–399
- Grapputo, A., S. Boman, L. Lindstrom, A. Lyytinen and J. Mappes, 2005. The voyage of an invasive species across continents: genetic diversity of North American and European Colorado potato beetle populations. *Mol. Ecol.*, 14: 4207–4219
- Gunderina, L.I., I.I. Kiknadze, A.G. Istomina and M. Batler, 2009. Geographic differentiation of genomic DNA of Chironomus plumosus (Diptera: Chironomidae) in natural holarctic populations. *Genetika*, 45: 64–72
- Hadrys, H., M. Balick and B. Schierwater, 1992. Applications of random amplified polymorphic DNA (RAPD) in molecular ecology. *Mol. Ecol.*, 1: 55–63
- Haymer, D.S. and D.O. McInnis, 1994. Resolution of populations of the Mediterranean fruit fly at the DNA level using random primers for the polymerase chain reaction. *Genome*, 37: 244–248
- Hiragi, C., K. Simões, E. Martins, P. Queiroz, L. Lima and R. Monnerat, 2009. Genetic variability in *Aedes aegypti* (L.) (Diptera: Culicidae) populations using RAPD markers. *Neotrop. Entomol.*, 38: 542–547
- Kaakeh, W., 2005. Longevity, fecundity, and fertility of the red palm weevil, *Rhynchophorus ferrugineus* Olivier (Coleoptera: Curculionidae) on natural and artificial diets. *Emirat J. Agric. Sci.*, 17: 23–33
- Karut, K. and C. Kazak, 2005. Akdeniz Bölgesi'nde yeni bir Hurma agaci (*Phoenix dactylifera* L.) zararlisi: *Rhynchophorus ferrugineus* (Olivier, 1790) (Coleoptera: Curculionidae). *Türk Entomol. Derg.*, 29: 295–300
- Kehat, M., 1999. Threat to Date Palms in Israel, Jordan and the Palestinian Authority by the Red Palm Weevil, *Rhynchophorus ferrugineus*. *Phytoparasitica*, 27: 107–108
- Kim K.S. and T.W. Sappington, 2004. Boll weevil (Anthonomus grandis Boheman) (Coleoptera: Curculionidae) dispersal in the southern United States: Evidence from mitochondrial DNA variation. Environ. Entomol., 33: 457–470
- Kontodimas, D.C., P. Milonas, V. Vassiliou, N. Thymakis and D. Economou, 2007. The occurrence of *Rhynchophorus ferrugineus* in Greece and Cyprus and the risk against the native Greek palm tree *Phoenix theofrasti. Entomologia Hellenica*, 16: 11–15
- Lala, E.R.P., M.H. Andó, L. Zalloum, M. Bértoli, M. de Oliveira Machado Dalalio, T.G.V. Silveira, M.L. Gomes, T.A. Guedes and S.M. De Araújoe, 2009. *Trypanosoma cruzi*: Different methods of data analysis to evaluate the genetics–biology relationship. *Exp. Parasitol.*, 123: 173–181
- Marimuthu, M., Y. Perumal, A.P. Salim and G. Sharma, 2009. Genetic similarity of eggplant shoot and fruit borer, *Leucinodes orbonalis*, populations. *DNA Cell Biol.*, 28: 599–603
- Ministry for Resources and Rural Affairs of Malta, 2008. Annual Reports of Government Departments. Available at: http://www.agric.gov.mt/file.aspx?f=410
- Murphy, S.T. and B.R. Briscoe, 1999. The red palm weevil as an alien invasive: biology and the prospects for biological control as a component of IPM. *Biocont. News Informa.*, 20: 35N–46N

- Nei, M. and W.H. Li, 1979. Mathematical model for studying genetic variation in terms of restriction endonucleases. P. Natl. Acad. Sci. USA, 76: 5269–5273
- Nirula, K.K., 1956. Investigations on the pests of coconut palm. Part IV. Rhynchophorus ferrugineus F. Indian Coconut J., 9: 229–247
- Perring, T.M., C.A. Farrar, A.D. Cooper and T.S. Bellows, 1993. Determining whitefly species. *Science*, 261: 1333–1335
- Sadakathulla, S., 1991. Management of Red Palm Weevil Rhynchophorus ferrugineus F. in Coconut Plantations. Planter, 67: 415–419
- Salama, H.S. and M.M. Saker, 2002. DNA fingerprints of three different forms of the red palm weevil collected from Egyptian date palm orchards. Arch. Phytopathol. Plant Prot., 35: 299–306
- Sharma, A.K., M.J. Mendki, S.N. Tikar, K. Chandel, D. Sukumaran, B.D. Parashar, V. Veer and O.P.S. Agarwaland Prakash, 2009. Genetic variability in geographical populations of *Culex quinquefasciatus* Say (Diptera: Culicidae) from India based on random amplified polymorphic DNA analysis. *Acta Trop.*, 112: 71–76
- Sivapragasam, A., A. Arikiah and C.A. Ranjit, 1990. The Red Stripe Weevil, *Rhynchophorus schach* Olivier (Coleoptera: Curculionidae): an Increasing Menace to Coconut Palms in Hilir Perak. *Planter*, 66: 113–123

- Sneath, P.H.A. and R.R. Sokal, 1973. Numerical Taxonomy: the Principles and Practice of Numerical Classification, p: 573. W.H. Freeman, San Francisco, California
- Srivastava, P.P., K. Vijayan, A.K. Awasthi, P.K. Kar, K. Thangavelu and B. Saratchandra 2005. Genetic analysis of silkworms *Bombyx mori* through RAPD markers. *Indian J. Biotechnol.*, 4: 389–395
- Tamura, K., J. Dudley, M. Nei and S. Kumar, 2007. MEGA4: Molecular evolutionary genetics analysis (MEGA) software version 4.0. *Mol. Biol. Evol.*, 24: 1596–1599
- Vieira, P., W. Burgermeister, M. Mota, M. Kai and G. Silva, 2007. Lack of genetic variation of *Bursaphelenchus xylophilus* in Portugal revealed by RAPD-PCR analyses. J. Nematol., 39: 118–126
- Wattanapongsiri, A., 1966. A Revision of the Genera Rhynchophorus and Dynamis (Coleoptera: Curculionidae). Dept. Agric. Sci. Bull., 1: 1– 328
- Williams, J.G.K., A.R. Kubelik, K.J. Livak, J.A. Rafalski and S.V. Tingey, 1990. DNA polymorphisms amplified by arbitrary primers as useful as genetic markers. *Nucl. Acid Res.*, 18: 6531–6535

(Received 22 June 2010; Accepted 01 August 2010)